IN VIVO SC & DRG ELECTROPHYSIOLOGY

PATCH CLAMP

# Characterization of Human iPSC-derived Neurons



## Axol sensory neurons plated at low density on astrocytes are improved



- 1. Axol sensory neurons plated at low density on astrocytes grew for >5 weeks without "balling up".
- 2. Single cells could be readily recorded by patch clamp electrophysiology.



### **Tetracaine blocks Na currents in a voltage-dependent fashion**



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Axol hiPSC sensory neuron action potentials faithfully follow stimulation



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**Tetracaine block of action potentials is frequency-dependent** 

100 action potentials elicited at each test concentration at each frequency





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## **Tetracaine block of action potentials is frequency-dependent**



## **Tetracaine block of action potentials is frequency-dependent**



No effect on resting membrane potential

Reduced excitability at 10 Hz > 3 Hz > 1 Hz seen in 2 endpoints

- 1. Number of action potentials >+20 mV
- 2. Peak amplitude of action potentials



## Retigabine hyperpolarized the resting membrane potential and decreased action potential firing in human iPSC-derived sensory neurons





## **Retigabine hyperpolarized the resting membrane potential in human iPSC-derived sensory neurons**





### **Retigabine hyperpolarized the resting membrane potential in human iPSC-derived sensory neurons**



	baseline	retigabine	retigabine + XE-991	wash
n	21	21	19	19
average ± sem	-54.7 ± 0.9 mV	-64.7 ± 0.7 mV	-53.1 ± 1.7 mV	-53.3 ± 1.1 mV
p-value (paired t-test)		<0.0001 (vs baseline)	<0.0001 (vs retigabine)	



	baseline	vehicle	vehicle + XE-991	wash
n	21	21	16	20
average ± sem	-51.8 ± 1.2 mV	-52.5 ± 1.3 mV	-52.2 ± 1.9 mV	-50.7 ± 1.7 mV
p-value (paired t-test)		0.1411 (vs baseline)	0.1962 (vs vehicle)	



## **Retigabine decreased action potential firing in human iPSC-derived sensory neurons**







## **Retigabine decreased action potential firing in human iPSC-derived sensory neurons**

XE-991 reversed the effect of retigabine



#### XE-991 "reduced" APs in vehicle treated cells?





## Retigabine increased the rheobase (current to elicit 1<sup>st</sup> action potential) in human iPSC-derived sensory neurons



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	baseline	retigabine	retigabine + XE-991	wash
n	18	18	16	14
average ± sem	22.2 ± 4.5 pA	64.4 ± 7.1 pA	23.1 ± 4.0 pA	18.8 ± 3.6 pA
p-value (paired t-test)		<0.0001 (vs baseline)	<0.0001 (vs retigabine)	

	baseline	vehicle	vehicle + XE-991	wash
n	17	17	15	17
average ± sem	20.0 ± 3.2 pA	21.8 ± 4.0 pA	30.0 ± 5.8 pA	22.4 ± 4.3 pA
p-value (paired t-test)		0.1876 (vs baseline)	0.0262 (vs vehicle)	



## Retigabine decreased the number of action potentials elicited by 3X rheobase in human iPSC-derived sensory neurons



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	baseline	retigabine	retigabine + XE-991	wash
n	16	16	16	14
average ± sem	14.4 ± 1.6	1.6 ± 0.7	12.0 ± 1.6	14.7 ± 1.8
p-value (paired t-test)		<0.0001 (vs baseline)	<0.0001 (vs retigabine)	

	baseline	vehicle	vehicle + XE-991	wash
n	17	17	15	15
average ± sem	15.6 ± 1.3	14.7 ± 1.3	7.1 ± 1.8	9.7 ± 1.4
p-value (paired t-test)		0.0156 (vs baseline)	<0.0001 (vs vehicle)	



## **Summary / Conclusions**

- 1. Axol sensory neurons grow best when plated at low density on a monolayer of supportive astrocytes.
- 2. Axol sensory neurons are excitable (evoked action potentials) but exhibit little spontaneous activity (like human DRG sensory neurons).
- 3. Axol sensory neurons express sufficient TTX-S and TTX-R Na currents to support drug discovery of Nav1.7 and Nav1.8 inhibitors.
- 4. Axol sensory neurons were pharmacologically validated using:
  - a) TTX
  - b) tetracaine
  - c) retigabine
- 5. Human iPSC-derived sensory neurons from Axol reproduce many properties expected from bona fide DRG sensory neurons and may be useful for supporting drug discovery programs for pain.



### **Contact us**



#### Raymond Price, PhD, EMBA Chief Business Officer Raymond.price@neuroservices-alliance.com

Mobile - +1 (858) 649 9403

## **neuroservices**



#### **Bob Petroski, PhD** CSO, Cell Electrophysiology

Bob.petroski@neuroservices-alliance.com Mobile - +1 (858) 774 4485

## The CNS Electrophysiology CRO

WWW.NEUROSERVICES-ALLIANCE.COM

# **Annex** Additional Data

## **Cell Capacitance and Membrane Resistance**



	Cm	Rm
Hi density w/o astrocytes	26.1 ± 0.5 pF (n=188)	442 ± 28 MΩ (n=187)
Low density on astrocytes	29.9 ± 0.6 pF (n=370)	286 ± 13 MΩ (n=370)

- 1. Axol sensory neurons grew larger with time in culture.
  - Increased Cm

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- Decreased Rm
- 2. Axol sensory neurons growing at low density on astrocytes were slightly, but significantly larger that cells plated at high density without astrocytes.



## **Resting membrane potential (K internal)**



- 1. Axol sensory neurons exhibited a negative resting membrane potential (RMP) early after plating.
- 2. This suggests that the Na-K ATPase and K leak channels responsible for establishing the RMP are already expressed by neural precursors.
- 3. Further differentiation in culture did not result in more negative RMP.
- 4. There was no difference in Vm between cells plated at high density and cells plated at low density on astrocytes.



## Spontaneous and evoked action potentials (K internal)



	# spontaneous APs in 1 min	# evoked APs in 1 sec
Hi density w/o astrocytes	0.7 ± 0.5 (n=80)	12.0 ± 1.1 (n=106)
Low density on astrocytes	17.2 ± 4.1 (n=178)	16.6 ± 0.7 (n=185)

INCLUSION CRITERIA: Vm <-40 mV

- 1. Axol sensory neurons exhibited very little spontaneous action potential firing.
- 2. More spontaneous activity was observed in cells plated on astrocytes.
- 3. Depolarizing current injection for 1 sec elicited only 1 action potential in most neurons in the first week in vitro.
- 4. However, the maximum number of action potentials elicited increased with time in culture.



## **Spontaneous action potentials (K internal)**



	# cells NO spontaneous APs	# cells ≥1 spontaneous APs
Hi density w/o astrocytes	77	3
Low density on astrocytes	140	38

INCLUSION CRITERIA: Vm <-40 mV



- 1. Axol sensory neurons exhibited very little spontaneous action potential firing.
- 2. More spontaneous activity was observed in cells plated on astrocytes (21% vs 4%).
- 3. The cells that did exhibit spontaneous activity, fired APs at a high rate (average 81 spikes per min).



## Na currents and K currents (K internal)





	Na current peak	K current at +60 mV
Hi density w/o astrocytes	5,482 ± 283 pA (n=116)	4,480 ± 283 pA (n=116)
Low density on astrocytes	5,347 ± 195 pA (n=224)	4,078 ± 187 pA (n=224)

- 1. Axol sensory neurons exhibited large voltage gated Na and K currents.
- 2. Both Na current and K current amplitudes\s increased with time in culture.
- 3. There was no significant difference in the amplitudes of Na currents and K currents for cells plated at high density and cells plated at low density on astrocytes.



## Na currents (Cs internal)





	Total Na current at -20 mV	TTX-R current at -20 mV
Hi density w/o astrocytes	6,292 ± 284 pA (n=67)	775 ± 51 pA (n=67)
Low density on astrocytes	8,630 ± 336 pA (n=104)	1,696 ± 109 (n=104)

- 1. Axol sensory neurons exhibited large voltage gated Na currents.
- 2. 10-15% of the total Na current was resistant to TTX (0.5 uM).
- 3. Total and TTX-R Na currents amplitudes increased with time in culture.
- 4. Na and TTX-R Na currents amplitudes were larger in cells growing at low density on astrocytes.



## **Axol cells >20 DIV: Capacitance and membrane resistance**







	Cm	Rm	Vm
Hi density	30.2 ± 0.8 pF	335 ± 18 MΩ	-49.7 ± 0.9 mV
w/o astrocytes	(n=79)	(n=78)	(n=55)
Low density on astrocytes	37.5 ± 0.9 pF	190 ± 8 MΩ	-50.4 ± 0.8 mV
	(n=178)	(n=178)	(n=89)

- 1. Axol sensory neurons growing at low density on astrocytes were significantly larger that cells plated at high density without astrocytes.
- 2. There was no difference in Vm between cells plated at high density and cells plated at low density on astrocytes.



## Axol cells >20 DIV: Na currents and K currents (K internal)







- 1. Axol sensory neurons exhibited large voltage gated Na and K currents.
- 2. There was no significant difference in the amplitudes of Na currents and K currents for cells plated at high density and cells plated at low density on astrocytes.

