BRAIN SLICE ELECTROPHYSIOLOGY

IN VIVO BRAIN ELECTROPHYSIOLOGY

IN VIVO SC & DRG ELECTROPHYSIOLOGY

MULTI ELECTRODE ARRAY

Network oscillations



SUMMARY

Network oscillations

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INTRODUCTION

Brain rhythms are periodically fluctuating waves of neuronal activity. Such rhythms reflect the synchronized activity of large numbers of neurons because synchronous currents sum together to generate large-amplitude fluctuations.

Different types of brain rhythms are observed in humans, primates, as well as rodents and occur in many cortical and subcortical regions. Each type of rhythm is observed during particular behaviors, generated by specific mechanisms, associated with characteristic neuronal firing properties and are thought to perform distinct functions.

Different brain regions display various oscillatory activities, for instance slow oscillation (<1 Hz), delta (1-4 Hz), theta (4-12 Hz), beta (12-30 Hz), gamma oscillations (20-80Hz) and sharp waves ripples (150-250 Hz).

Within the hippocampus, theta, gamma oscillations and sharp wave ripples could be observed.

- Theta oscillations (4-12 Hz) arise during active exploration and rapid eye movement (REM) sleep
- gamma oscillations (20-80 Hz) are associated with cognitive function
- Sharpe wave ripples (150-250 Hz) are observed in animals during waking immobility and during slow wave sleep. Sharp waves ripples consist in a replay of previous activity in a temporally compressed manner, which is necessary for memory consolidation.

The most ubiquitous rhythms are gamma oscillations. This rhythms also represent a relevant model of investigation in CNS drug discovery as it is often impaired in disorders involving cognitive deficits, such as schizophrenia, Alzheimer's disease, and depression.



INTRODUCTION



(E) Excitatory, (I) inhibitory, (PSCs) postsynaptic currents (P) pyramidal cell, (B) basket interneuron (Buzsáki and Wang, 2012)

Gamma oscillations mechanisms

Gamma oscillations are generated by reciprocal connections between interneurons and pyramidal cells. Particular perisomatic-targeting interneurons - parvalbumin-positive basket cells - play a critical role in synchronizing the activity of numerous pyramidal cells by fast rhythmic inhibition (Bartos et al, 2007).

To summarize, excitatory cells provide feedback excitation to the circuit, while interneurons contribute phasic drive to pace the oscillations.



MATERIALS & METHODS

Hippocampal slices - CA3 recording (1/3)



Area of recording



MATERIALS & METHODS

Data analysis (2/3)





MATERIALS & METHODS

Data analysis (3/3)

Compound testing – paired comparison



Assessment of the power of oscillations

Phenotyping – unpaired comparison



Assessment of the predominant frequency of oscillations as a function of time







In vitro models (1/4)

Network oscillations in hippocampus

Gamma oscillations arise in the cortex as well as in the hippocampus. However, several features of the hippocampus facilitate the study of such oscillations.

- · Gamma oscillations are an emergent property of the circuitry in the hippocampus .
- The hippocampus contains densely packed neurons that generate large local field potentials and, in turn, generate large rhythms. This is especially true for CA3 and CA1 regions, because the pyramidal cell dendrites are aligned in parallel. Because of this, synaptic currents flow in the same direction and sum together to produce large amplitude signals that are easily detected in local field potential recordings.
- Much is known about hippocampal interneurons that are important for rhythm generation.
- Persistent gamma oscillations (up to 2hours of recording) can be induced *in vitro* in both rat and mouse. Either ionotropic glutamatergic (kainate model) or muscarinic acetylcholine (carbachol model) receptors activation allow for the circuit to oscillate without external inputs.



In vitro models (2/4)



Carbachol induced oscillations by mimicking cholinergic input to the hippocampus via the septum during exploratory behavior in vivo. Oscillations are generated by alterning cycles of AMPA receptor mediated recurrent excitation followed by feedback inhibition from perisomatic-targeting interneurons. (Traub et al., 2000, Mann et al., 2005)





In vitro models (3/4)



Time (min)

neuroservices

receptors activation to both excitatory and inhibitory neurons in the hippocampus (Clarke et al., 1997; Fisahn et al., 2004). Network oscillations are induced in both rat and mouse hippocampal slices with maximal effect reached at 100 nM concentration.

In vitro models (4/4)

Donepezil-induced oscillations



Donepezil – an acetylcholinesterase inhibitor approved for symptomatic treatment of patients with mild-moderate AD – is capable of inducing network oscillations. The power of donepezil-induced oscillations continuously increased and did not reach stability after one hour of 1 μ M donepezil exposure.





From In vitro to in vivo (1/2)

Mechanisms

 Gamma oscillations can be recorded both *in vivo* using Electro-encephalogram (EEG) or field potentials recordings and *in vitro* using extracellular recordings. Both *in vitro* models (carbachol and kainate) are based on the phasic GABAergic inhibition onto pyramidal cells to generate gamma oscillations. The genesis of gamma oscillation *in vivo* is also based on phasic GABAergic inhibition. Moreover, Basket cells fire action potentials that are phaselocked to gamma oscillations and play a critical role in the oscillations genesis both *in vivo* and *in vitro*. To summarize, gamma oscillations *in vivo* and *in vitro* mechanisms are quite similar and based on phasic GABAergic inhibition primarily mediated by parvalbumin-positive basket cells.

Temperature susceptibility

• While gamma oscillations *in vivo* display a peak frequency of around 30-40 Hz, it is slightly lower *in vitro* as the frequency is very susceptible to temperature (Lu et al., 2012). Since recording for a long period of time at physiological temperatures causes cell death, recording temperature is kept at 32° C. As the consequence the predominant frequency of gamma oscillations recorded *in vitro* is shifted to around 20 Hz.



From In vitro to in vivo (2/2)

Consistent results between in vivo and in vitro

- **NMDA antagonists:** in accordance with literature, ketamine and MK-801 increase the power of gamma oscillations induced by either kainate or carbachol (Neuroservice data). *In vivo*, ketamine and MK-801 increase the power of cortical oscillations in the gamma range in rodent and humans (Hakami et al., 2009; Kocsis et al., 2012; Hiyoshi et al., 2014, Sanacora et al., 2014).
- Antipsychotic drugs: In vitro, clozapine serotoninergic & dopaminergic antagonist and haloperidol – dopaminergic antagonist – decrease the power of oscillations induced by kainate (Neuroservice data). In vivo, clozapine and haloperidol depressed the power of cortical oscillations in the gamma range (Jones et al., 2012).
- **Neurotransmitters**: 5-HT reduces gamma oscillations recorded *in vitro* (Neuroservice data) and *in vivo* in the prefrontal cortex (Puig, 2010). Norepinephrine also decreases gamma oscillations recorded *in vitro* (Neuroservice data) and *in vivo* (Brown et al., 2005) in the hippocampus.



RESULTS



RESULTS

AMPA/Kainate and GABA_A receptors

Summary Materials and methods

neuroservices

Picrotoxin, NBQX, diazepam

- Picrotoxin strongly inhibited both carbachol- and kainate-induced oscillations indicating a requirement of GABA_Amediated inhibition within the neuronal network to observe oscillations. NBQX strongly inhibited both carbachol- and kainate-induced oscillations indicating the importance of AMPA/Kainate mediated transmission in this process.
- 10 µM diazepam application reduced the frequency of network oscillations. The power of network oscillations was not strongly modified. Power



amplitude

Normalized



12

Control

0.1 μM

1 μM

10 µM



NMDA receptors(1/2)

Summary Materials and methods





NMDA receptors (2/2)



 The MK-801 – a NMDA receptor antagonist – slightly increased the power of network oscillations In slices exposed to CIQ – a potentiator of NMDA receptors containing GluN2C/GluN2D - the power and the frequency of oscillations was not modified.





Sodium channels





- --- Vehicle (6 slices from 2 rats)
- Carbamazepine (6 slices from 2 rats)

 Carbamazepine – a sodium channel inhibitor – did not modulate both the power and the frequency of carbacholinduced network oscillations when applied at 6 µM.



REFERENCE DATA

Dopaminergic receptors



Summary Materials and methods

Haloperidol PD-168077

Haloperidol – a dopaminergic receptor antagonist – applied at 10 μ M concentration decreased the power and the frequency of oscillations.

PD-168077 dopaminergic – a receptor D4 agonist - applied at 100 nM concentration did not modify the the power and frequency of network oscillations. As expected, ketamine applied at 100 µM increased the power and decreased frequency the of network oscillations.



RESULTS

Antipsychotic



 Clozapine – a serotonin and dopamine receptors antagonist used as antipsychotic agent – applied at 30 µM concentration decreased the power of network oscillations.





Serotoninergic receptors



5-HT dose-dependently decreased the power of network oscillations. Similarly, mCPBG – a 5-HT₃ agonist – applied at 100 μM concentration decreases the power and the frequency of oscillations.





Adrenergic receptors (1/2)

Summary Materials and methods

neuroservices



 Norepinephrine (NE) decreased the power and increased the frequency of kainate-induced network oscillations in both rat and mouse hippocampal slices.



Adrenergic receptors (2/2)



Summary Materials and methods

In the presence of an Adrenergic receptors blocker, norepinephrine did not substantially modulate kainate-induced network oscillations in rat hippocampal slices.

In kainate-induced network oscillations from α 1A KO mice, the effect of NE was reduced in comparison with the one in WT mice.





Somatostatin receptors

L803,087





 L-803,087 – a potent and selective SST4 agonis t– decreased both the power and the frequency of network oscillations when applied at 30 µM.





Cholinergic receptors (1/2)



Summary Materials and methods

Scopolamine – a non-selective muscarinic antagonist – blocked the network oscillations induced by carbachol but did not modify oscillations induced by kainate.





Cholinergic receptors (2/2)

Summary Materials and methods



 Anatabine – a cholinergic agonist – did not modulated the power of carbachol-induced oscillations but did slightly reduced the frequency.





Opioid receptors





 Fentanyl – a μ-opioid agonist – applied at 1 μM modulated carbachol-induced network oscillations in rat hippocampal slices by transiently increasing their power and decreasing their frequency.





Neuregulin-ERBb4 pathway





→ NRG-1 (5 slices from 3 rats)

 NRG-1β – an epidermal growth factor receptor (EGFR) agonist – increased the power and decreases the frequency of kainate-induced oscillations.





SK channels

TEA



Predominant Frequency



 The application of 0.5 mM Tetraethylammonium (TEA) – an inhibitor of Ca²⁺⁻activated K⁺ (SK) channels significantly increased the AUC of network oscillations and decreased their frequency.



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