MULTI ELECTRODE ARRAY

Network oscillations

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SUMMARY

Network oscillations

- Information about network oscillations
- Materials & Methods - [Go](#)
- *In vitro* Models - [Go](#)
- From *in vitro* to *in vivo* - [Go](#)

Reference data

- AMPA/Kainate and GABA$_A$ receptors – [Picrotoxine, NBQX](#)
- NMDA receptors – [Ketamine](#), [MK-801, CIQ](#)
- Sodium channels – [Carbamazepine](#)
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INTRODUCTION

Brain rhythms are periodically fluctuating waves of neuronal activity. Such rhythms reflect the synchronized activity of large numbers of neurons because synchronous currents sum together to generate large-amplitude fluctuations.

Different types of brain rhythms are observed in humans, primates, as well as rodents and occur in many cortical and subcortical regions. Each type of rhythm is observed during particular behaviors, generated by specific mechanisms, associated with characteristic neuronal firing properties and are thought to perform distinct functions.

Different brain regions display various oscillatory activities, for instance slow oscillation (<1 Hz), delta (1-4 Hz), theta (4-12 Hz), beta (12-30 Hz), gamma oscillations (20-80Hz) and sharp waves ripples (150-250 Hz).

Within the hippocampus, theta, gamma oscillations and sharp wave ripples could be observed.

- Theta oscillations (4-12 Hz) arise during active exploration and rapid eye movement (REM) sleep
- gamma oscillations (20-80 Hz) are associated with cognitive function
- Sharpe wave ripples (150-250 Hz) are observed in animals during waking immobility and during slow wave sleep. Sharp waves ripples consist in a replay of previous activity in a temporally compressed manner, which is necessary for memory consolidation.

The most ubiquitous rhythms are gamma oscillations. This rhythms also represent a relevant model of investigation in CNS drug discovery as it is often impaired in disorders involving cognitive deficits, such as schizophrenia, Alzheimer's disease, and depression.

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INTRODUCTION

Extracellular recording of local field potential
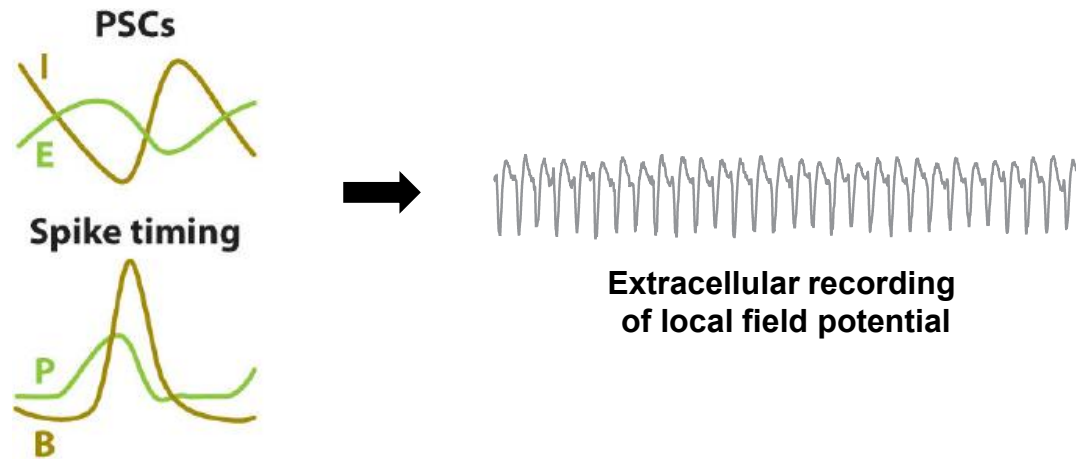
(E) Excitatory, (I) inhibitory, (PSCs) postsynaptic currents (P) pyramidal cell, (B) basket interneuron (Buzsáki and Wang, 2012)

Gamma oscillations mechanisms

Gamma oscillations are generated by reciprocal connections between interneurons and pyramidal cells. Particular perisomatic-targeting interneurons - parvalbumin-positive basket cells - play a critical role in synchronizing the activity of numerous pyramidal cells by fast rhythmic inhibition (Bartos et al, 2007).

To summarize, excitatory cells provide feedback excitation to the circuit, while interneurons contribute phasic drive to pace the oscillations.

Hippocampal slices – CA3 recording (1/3)
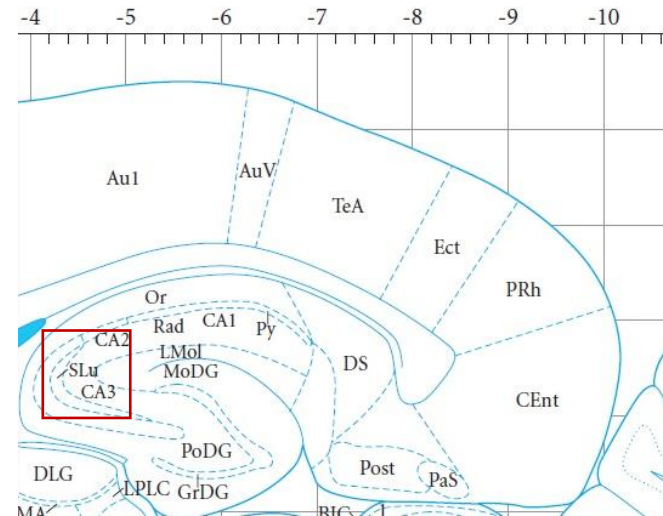
Area of recording

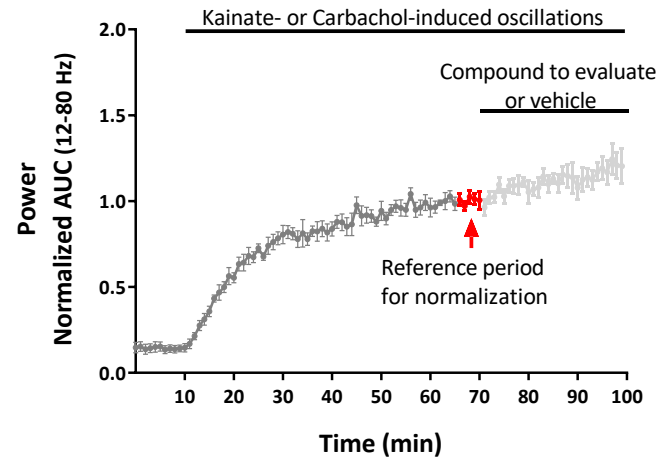**D a t a   a n a l y s i s   (2 / 3)**
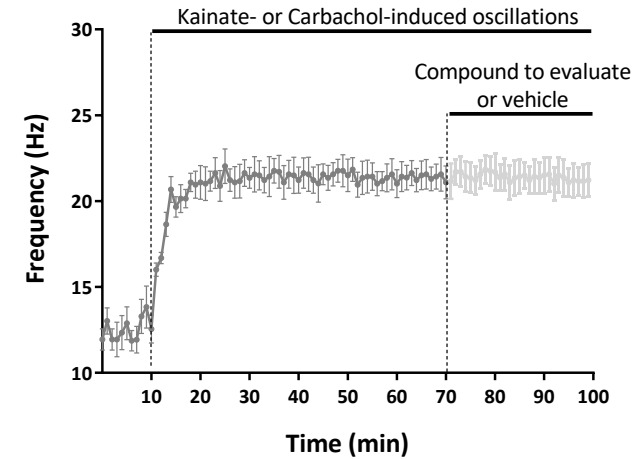
MATERIALS & METHODS

Data analysis (3/3)

**Compound testing –
paired comparison**

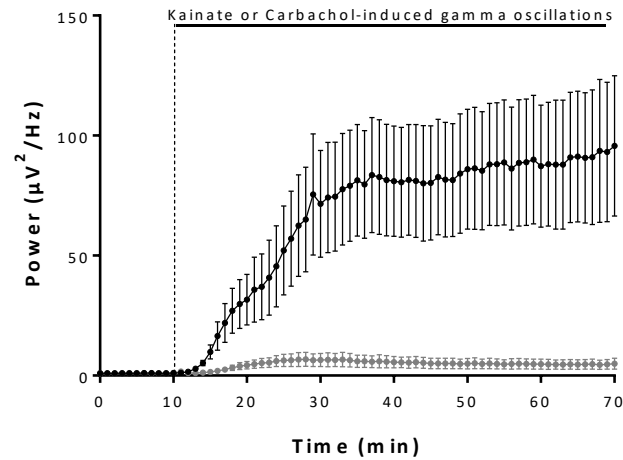### Assessment of the power of oscillations as a function of time

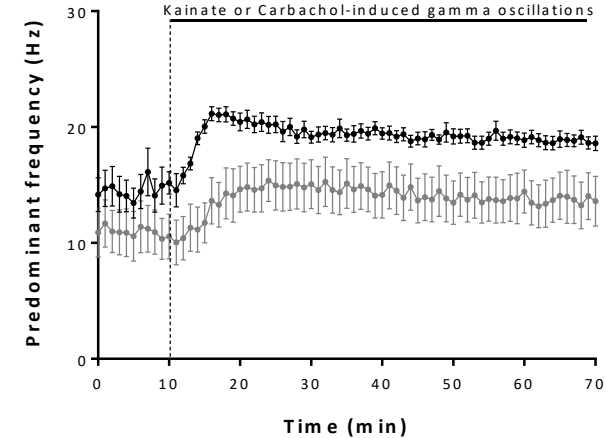### Assessment of the predominant frequency of oscillations as a function of time

**Phenotyping –
unpaired comparison**

Assessment of the power of oscillations as a function of time

Assessment of the predominant frequency of oscillations as a function of time

NETWORK OSCILLATIONS

In vitro models (1/4)

Network oscillations in hippocampus

Gamma oscillations arise in the cortex as well as in the hippocampus. However, several features of the hippocampus facilitate the study of such oscillations.
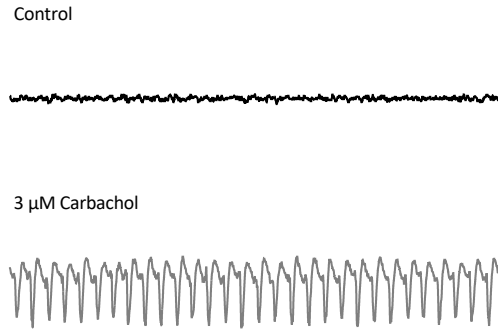
- Gamma oscillations are an emergent property of the circuitry in the hippocampus .

- The hippocampus contains densely packed neurons that generate large local field potentials and, in turn, generate large rhythms. This is especially true for CA3 and CA1 regions, because the pyramidal cell dendrites are aligned in parallel. Because of this, synaptic currents flow in the same direction and sum together to produce large amplitude signals that are easily detected in local field potential recordings.

- Much is known about hippocampal interneurons that are important for rhythm generation.

- Persistent gamma oscillations (up to 2hours of recording) can be induced *in vitro* in both rat and mouse. Either ionotropic glutamatergic (kainate model) or muscarinic acetylcholine (carbachol model) receptors activation allow for the circuit to oscillate without external inputs.
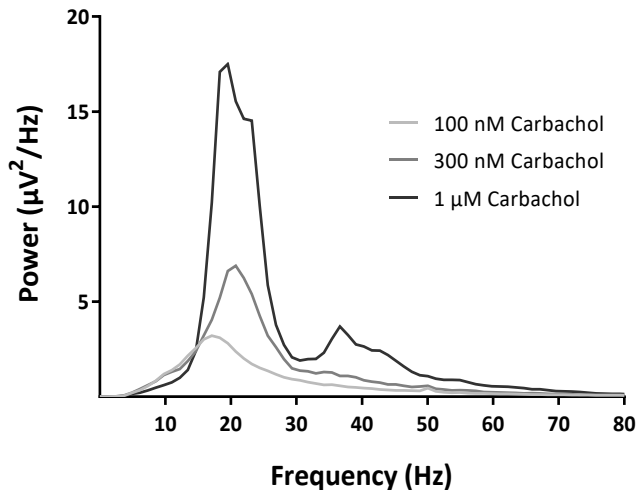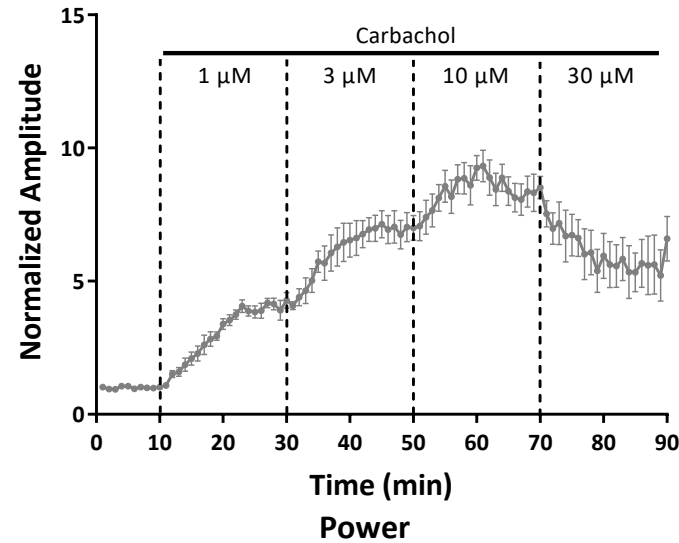
neuroservices
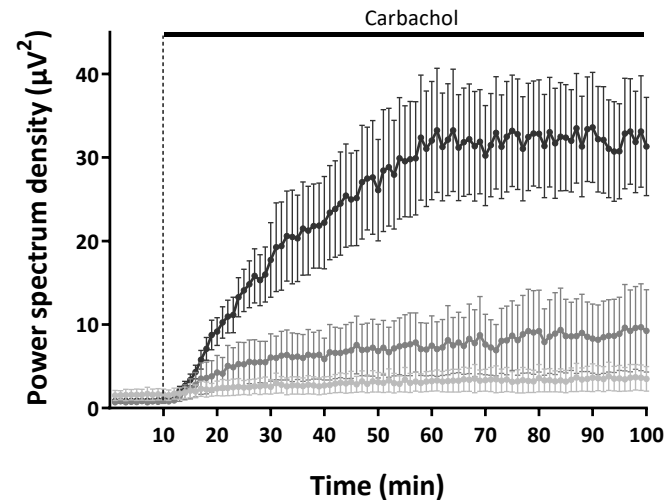ALLIANCE

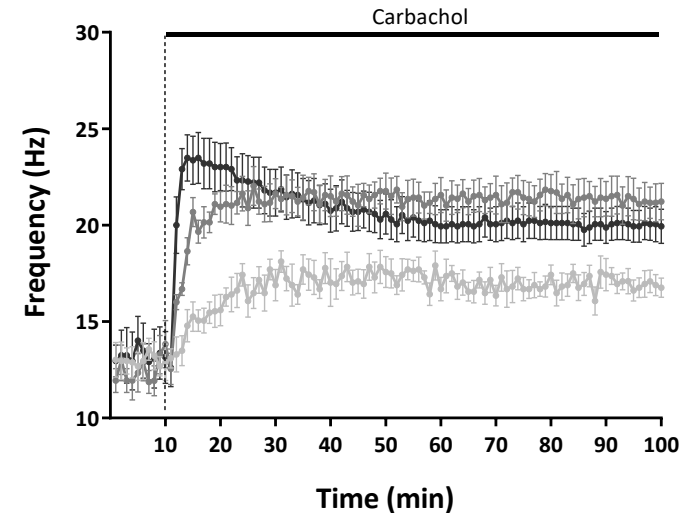## Carbachol-induced oscillations

Control

3 µM Carbachol

Carbachol dose-range (rat hippocampal slices)

Carbachol induced oscillations by mimicking cholinergic input to the hippocampus via the septum during exploratory behavior *in vivo*. Oscillations are generated by alternating cycles of AMPA receptor mediated recurrent excitation followed by feedback inhibition from perisomatic-targeting interneurons. (Traub et al., 2000, Mann et al., 2005)

Power spectra
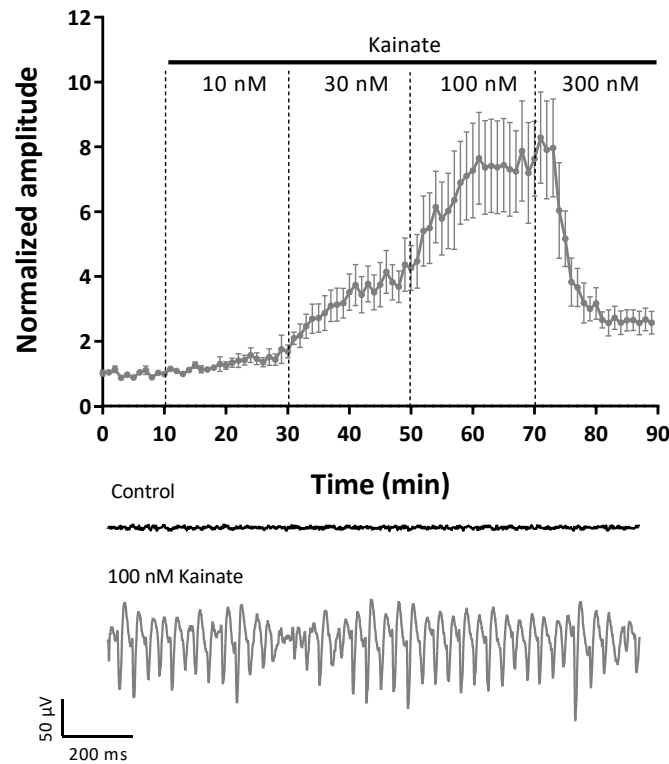

- 100 nM Carbachol
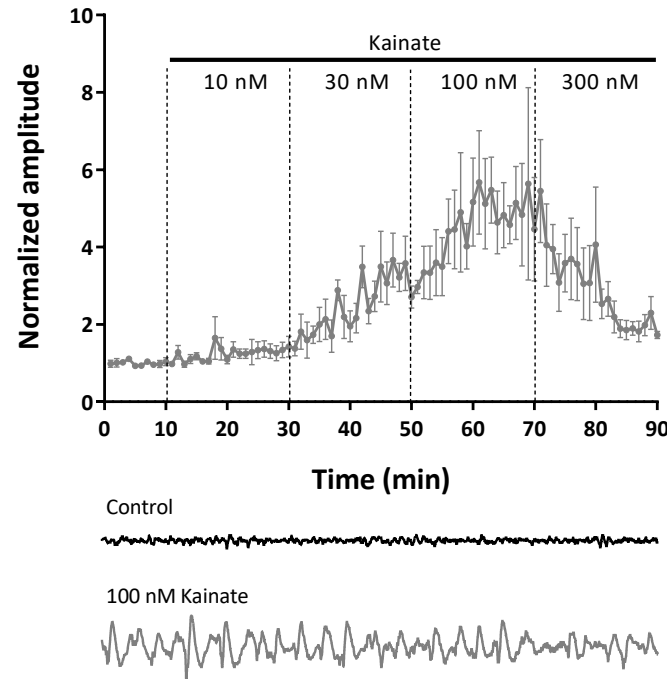- 300 nM Carbachol
- 1 µM Carbachol

Power

Frequency

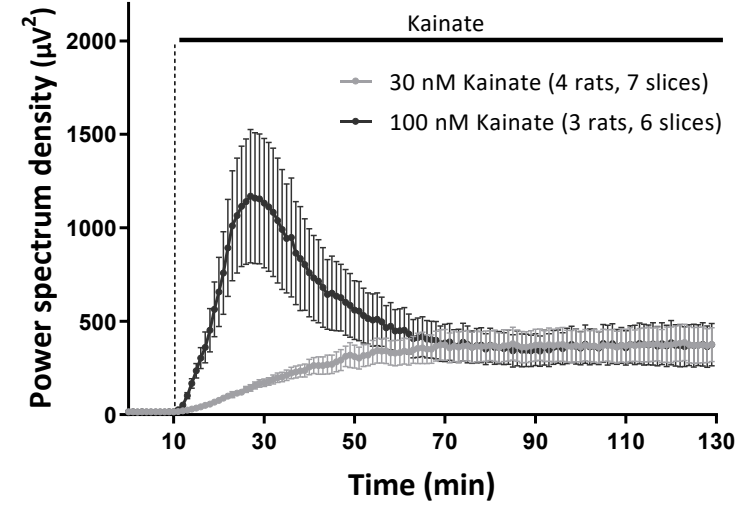

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Kainate-induced oscillations
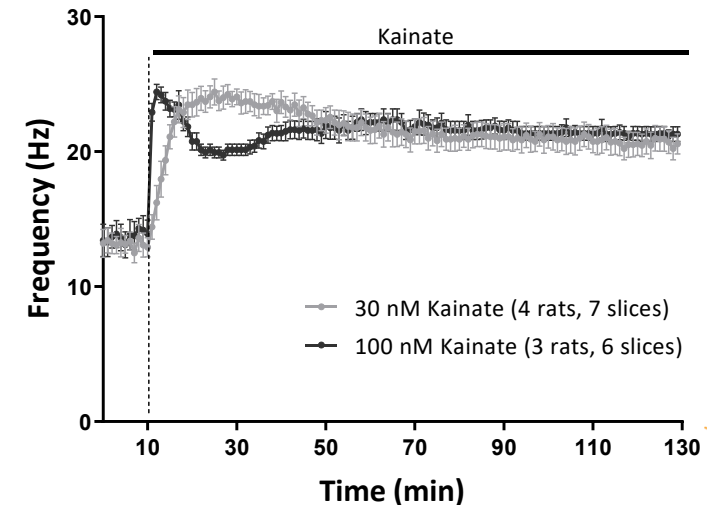
Kainate dose-range (rat hippocampal slices)

Kainate dose-range (mouse hippocampal slices)
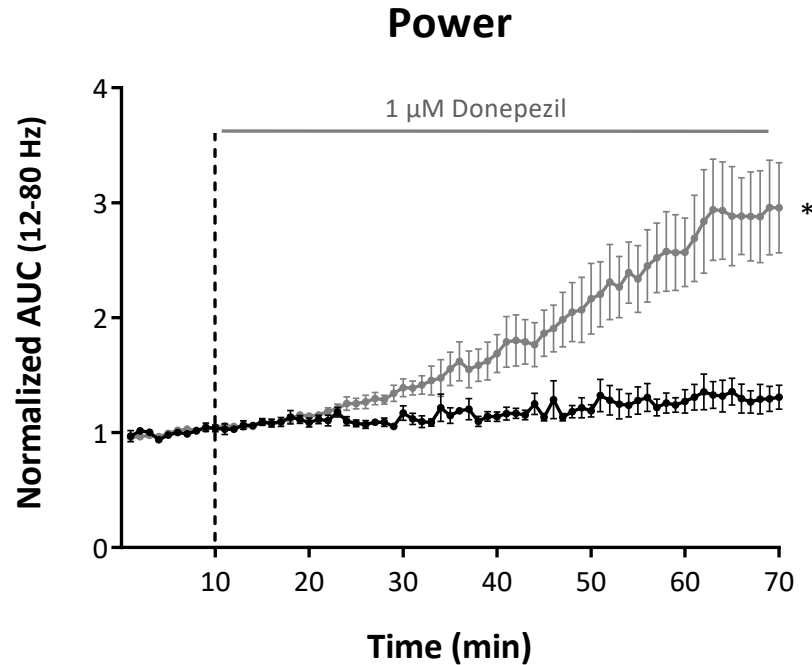
Power

Predominant Frequency

Kainate induces oscillations by providing tonic excitation via ionotropic glutamate receptors activation to both excitatory and inhibitory neurons in the hippocampus (Clarke et al., 1997; Fisahn et al., 2004). Network oscillations are induced in both rat and mouse hippocampal slices with maximal effect reached at 100 nM concentration.
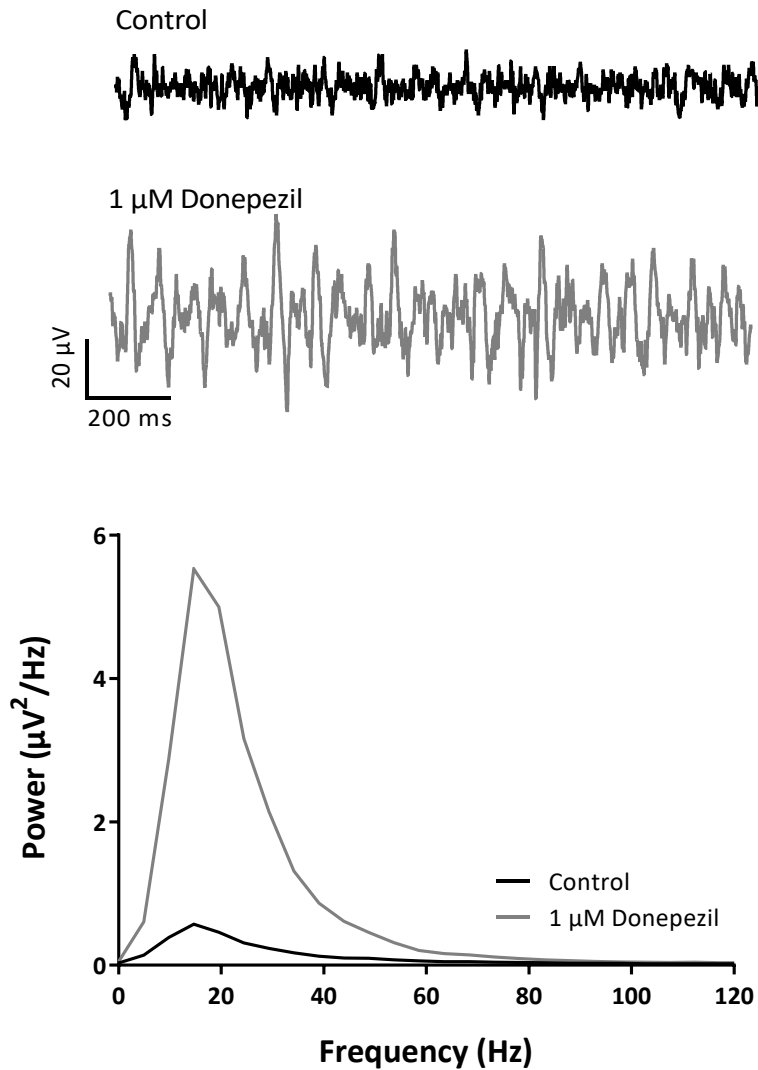
Donepezil-induced oscillations

Power

Donepezil – an acetylcholinesterase inhibitor approved for symptomatic treatment of patients with mild-moderate AD – is capable of inducing network oscillations. The power of donepezil-induced oscillations continuously increased and did not reach stability after one hour of 1 μM donepezil exposure.

Mechanisms

- Gamma oscillations can be recorded both *in vivo* using Electro-encephalogram (EEG) or field potentials recordings and *in vitro* using extracellular recordings. Both *in vitro* models (carbachol and kainate) are based on the phasic GABAergic inhibition onto pyramidal cells to generate gamma oscillations. The genesis of gamma oscillation *in vivo* is also based on phasic GABAergic inhibition. Moreover, Basket cells fire action potentials that are phase-locked to gamma oscillations and play a critical role in the oscillations genesis both *in vivo* and *in vitro*. To summarize, gamma oscillations *in vivo* and *in vitro* mechanisms are quite similar and based on phasic GABAergic inhibition primarily mediated by parvalbumin-positive basket cells.

Temperature susceptibility

- While gamma oscillations *in vivo* display a peak frequency of around 30-40 Hz, it is slightly lower *in vitro* as the frequency is very susceptible to temperature (Lu et al., 2012). Since recording for a long period of time at physiological temperatures causes cell death, recording temperature is kept at 32° C. As the consequence the predominant frequency of gamma oscillations recorded *in vitro* is shifted to around 20 Hz.

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Consistent results between *in vivo* and *in vitro*

- **NMDA antagonists:** in accordance with literature, ketamine and MK-801 increase the power of gamma oscillations induced by either kainate or carbachol (Neuroservice data). *In vivo*, ketamine and MK-801 increase the power of cortical oscillations in the gamma range in rodent and humans (Hakami et al., 2009 ; Kocsis et al., 2012 ; Hiyoshi et al., 2014, Sanacora et al., 2014).

- **Antipsychotic drugs:** *In vitro*, clozapine – serotoninergic & dopaminergic antagonist – and haloperidol – dopaminergic antagonist – decrease the power of oscillations induced by kainate (Neuroservice data). *In vivo*, clozapine and haloperidol depressed the power of cortical oscillations in the gamma range (Jones et al., 2012).

- **Neurotransmitters**: 5-HT reduces gamma oscillations recorded *in vitro* (Neuroservice data) and *in vivo* in the prefrontal cortex (Puig, 2010). Norepinephrine also decreases gamma oscillations recorded *in vitro* (Neuroservice data) and *in vivo* (Brown et al., 2005) in the hippocampus.

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RESULTS

RESULTS

AMPA/Kainate and GABA$_A$ receptors

Picrotoxin, NBQX, diazepam

- Picrotoxin strongly inhibited both carbachol- and kainate-induced oscillations indicating a requirement of GABA$_A$-mediated inhibition within the neuronal network to observe oscillations. NBQX strongly inhibited both carbachol- and kainate-induced oscillations indicating the importance of AMPA/Kainate mediated transmission in this process.

- 10 μM diazepam application reduced the frequency of network oscillations. The power of network oscillations was not strongly modified.
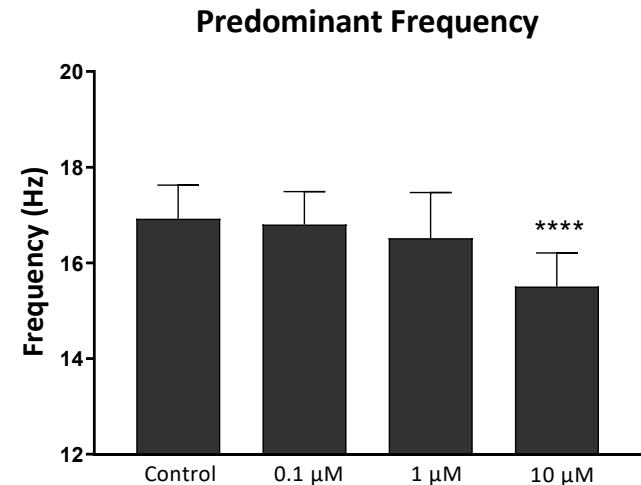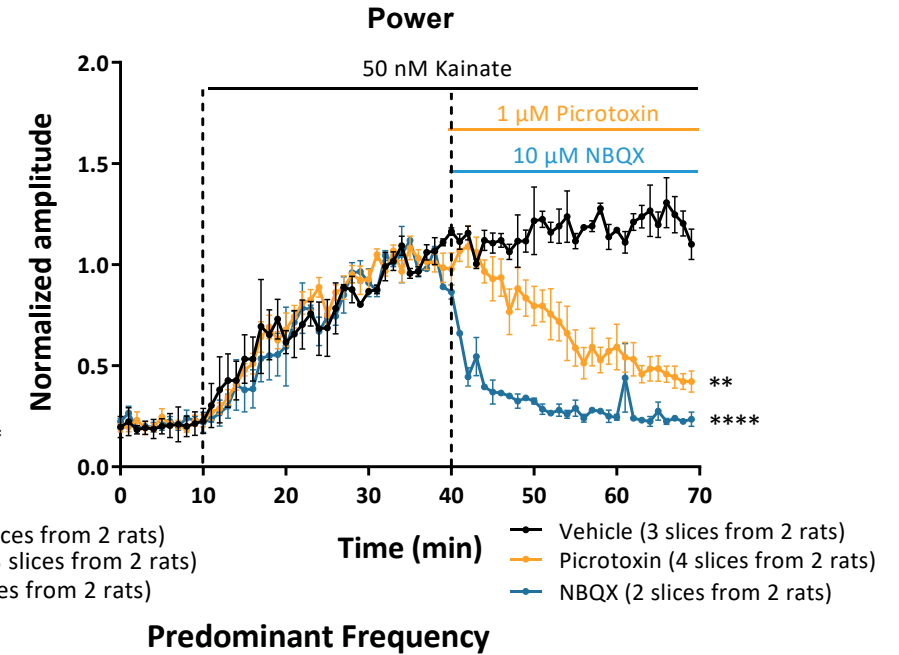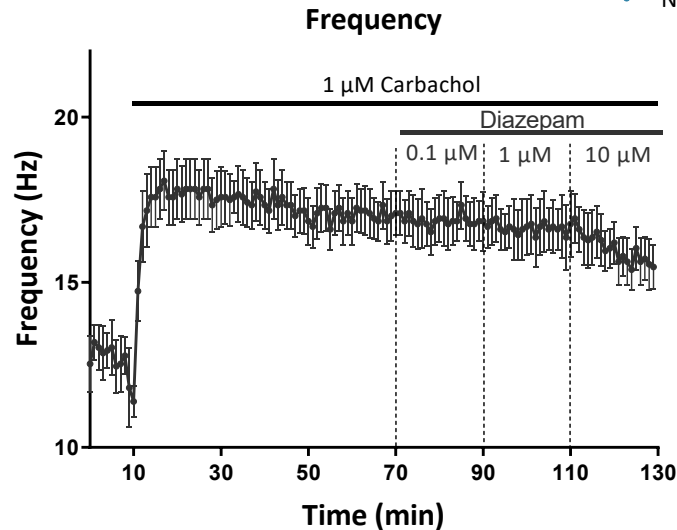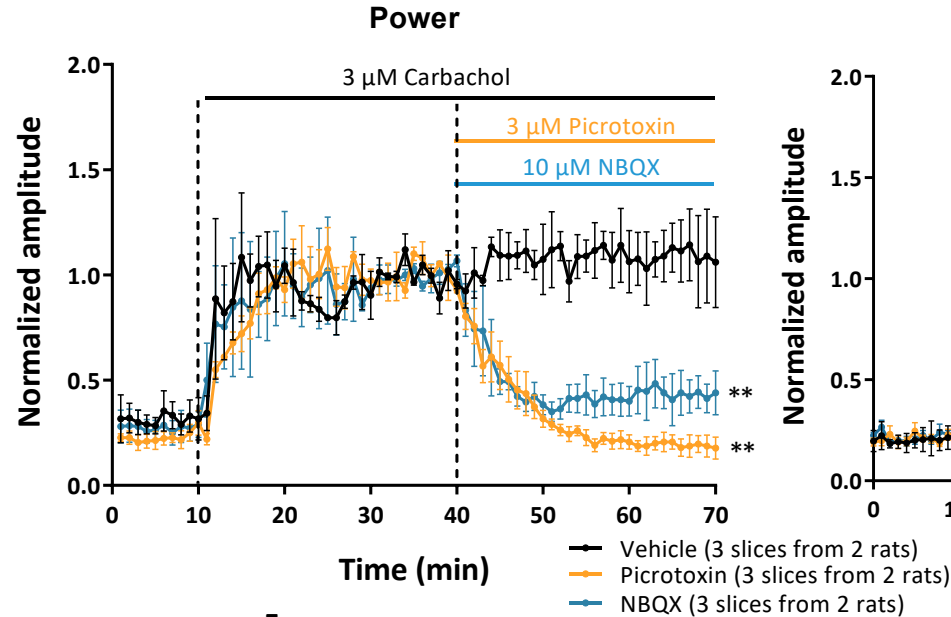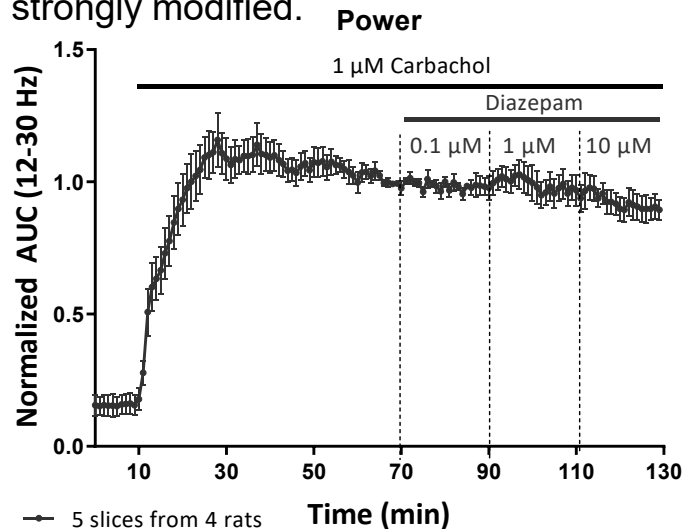
RESULTS

NMDA receptors(1/2)
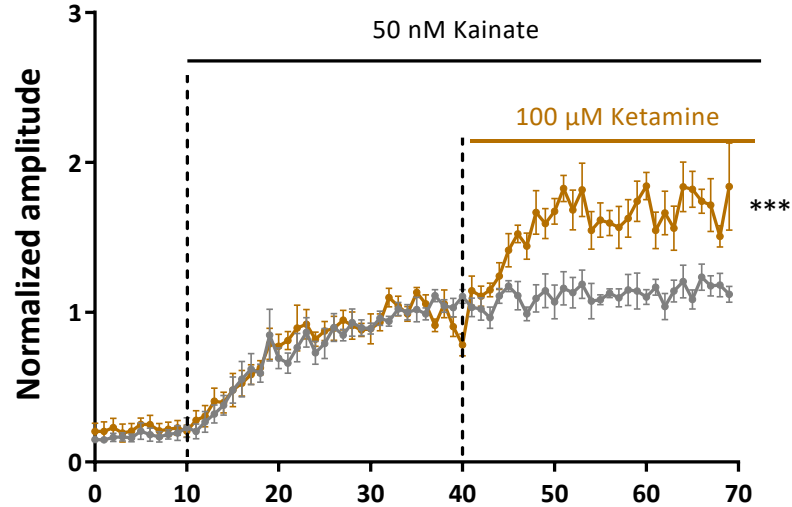
Ketamine

Power

- Ketamine – a NMDA receptor antagonist – applied at 100 µM concentration increased the power and decreased the frequency of kainate and carbachol-induced oscillations. From 500 µM concentration, ketamine abolished gamma oscillations.
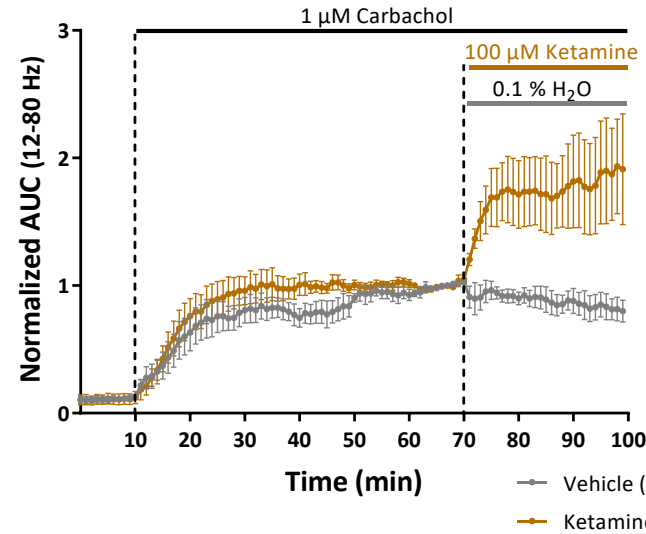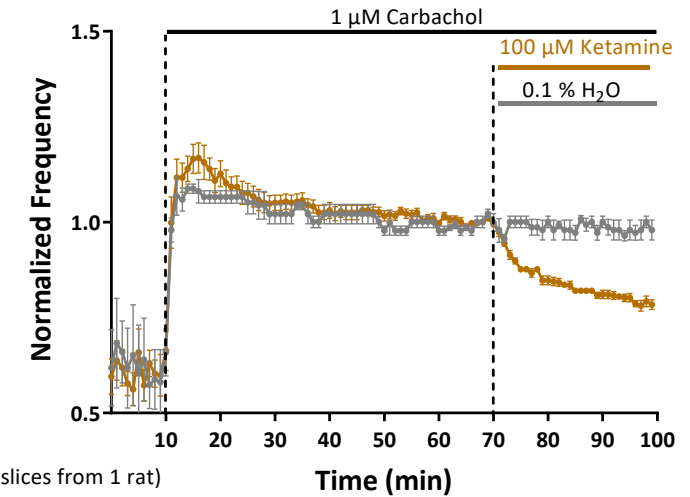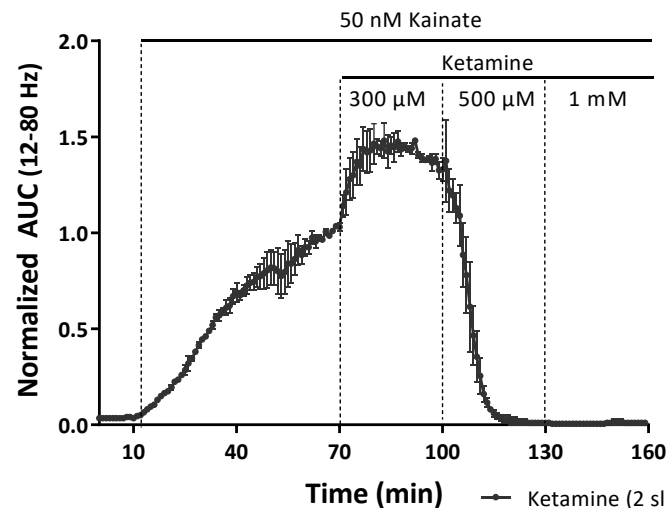
RESULTS

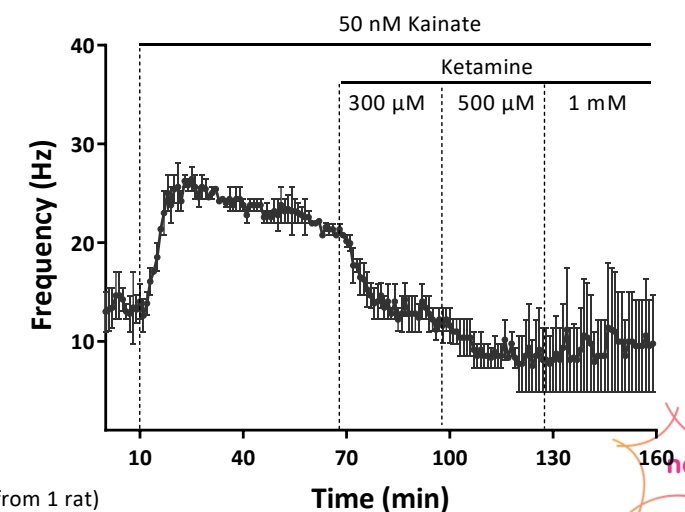## NMDA receptors (2/2)

MK-801, CIQ
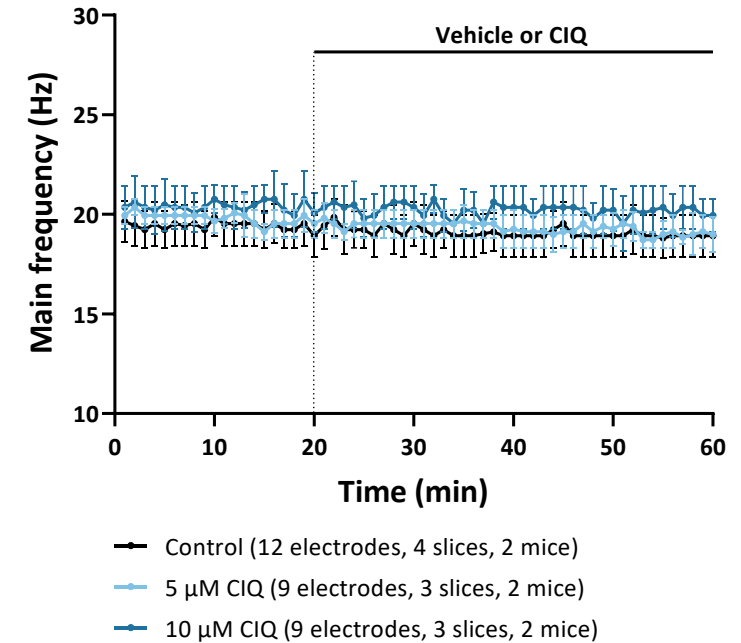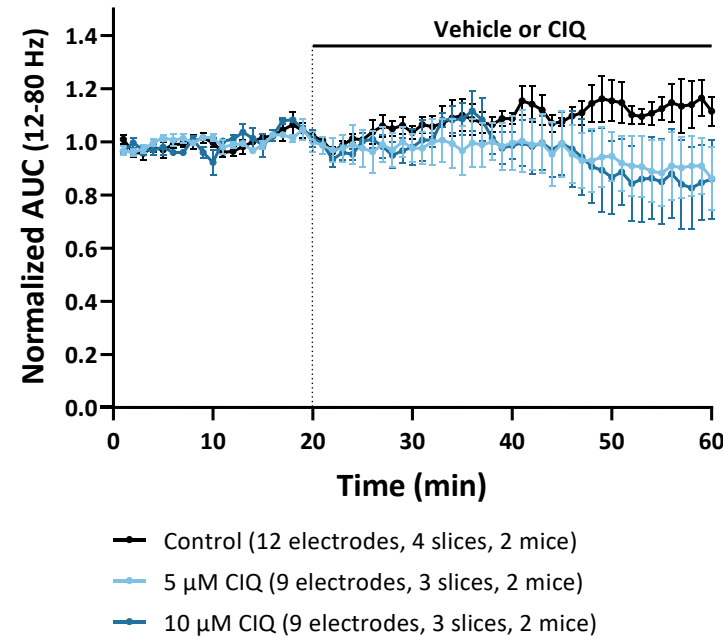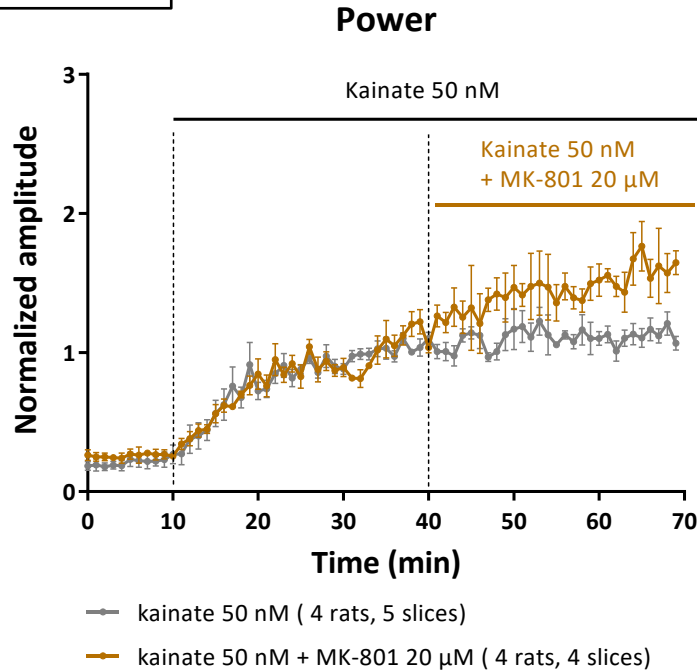
Power

- The MK-801 – a NMDA receptor antagonist – slightly increased the power of network oscillations

- In slices exposed to CIQ – a potentiator of NMDA receptors containing GluN2C/GluN2D - the power and the frequency of oscillations was not modified.
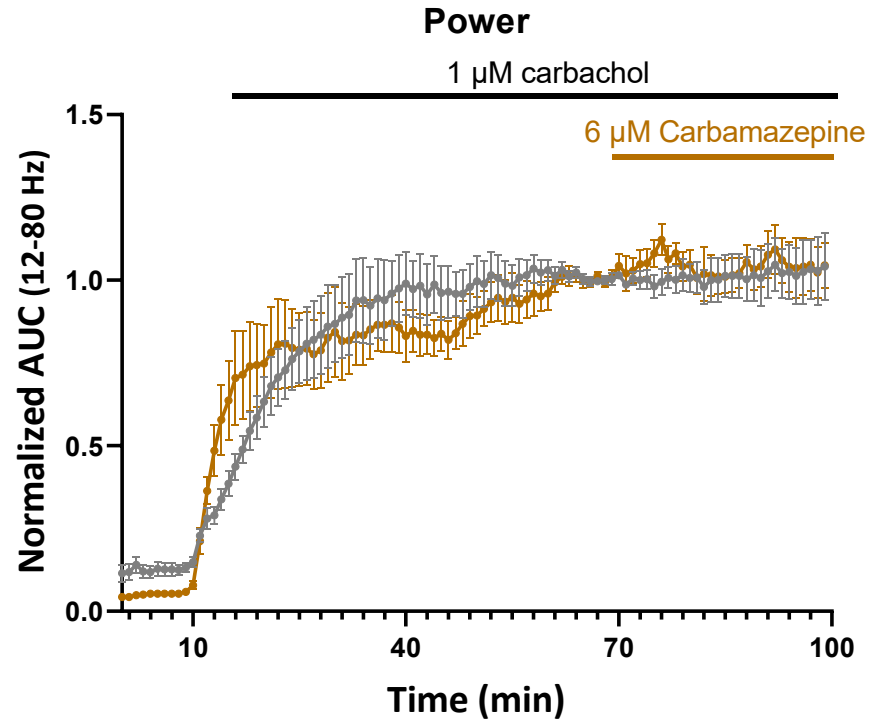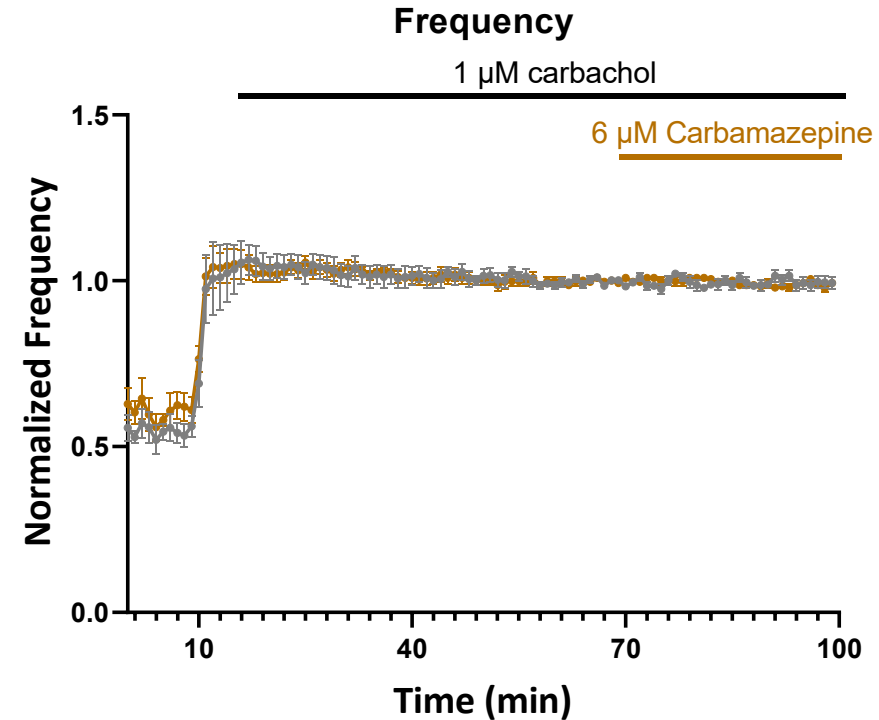
RESULTS

Sodium channels

Carbamazepine

Power

1 µM carbachol

6 µM Carbamazepine

Frequency

1 µM carbachol

6 µM Carbamazepine

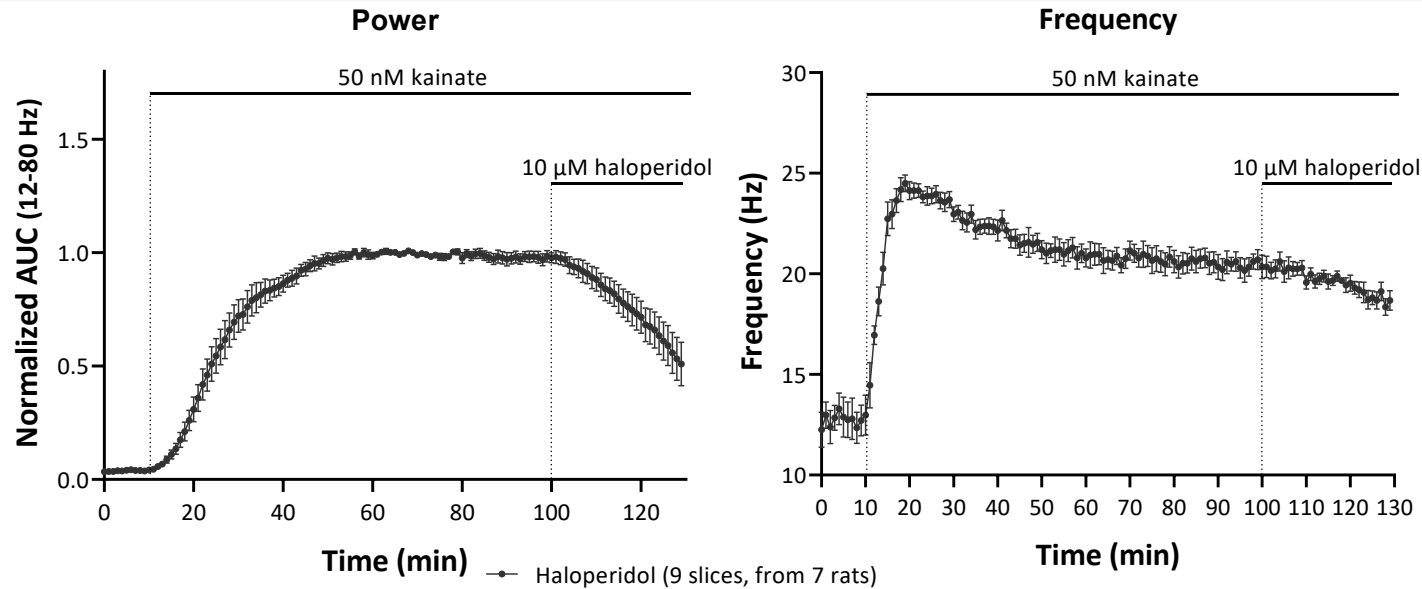— Vehicle (6 slices from 2 rats)
— Carbamazepine (6 slices from 2 rats)

- Carbamazepine – a sodium channel inhibitor – did not modulate both the power and the frequency of carbachol-induced network oscillations when applied at 6 µM.

REFERENCE DATA

Dopaminergic receptors

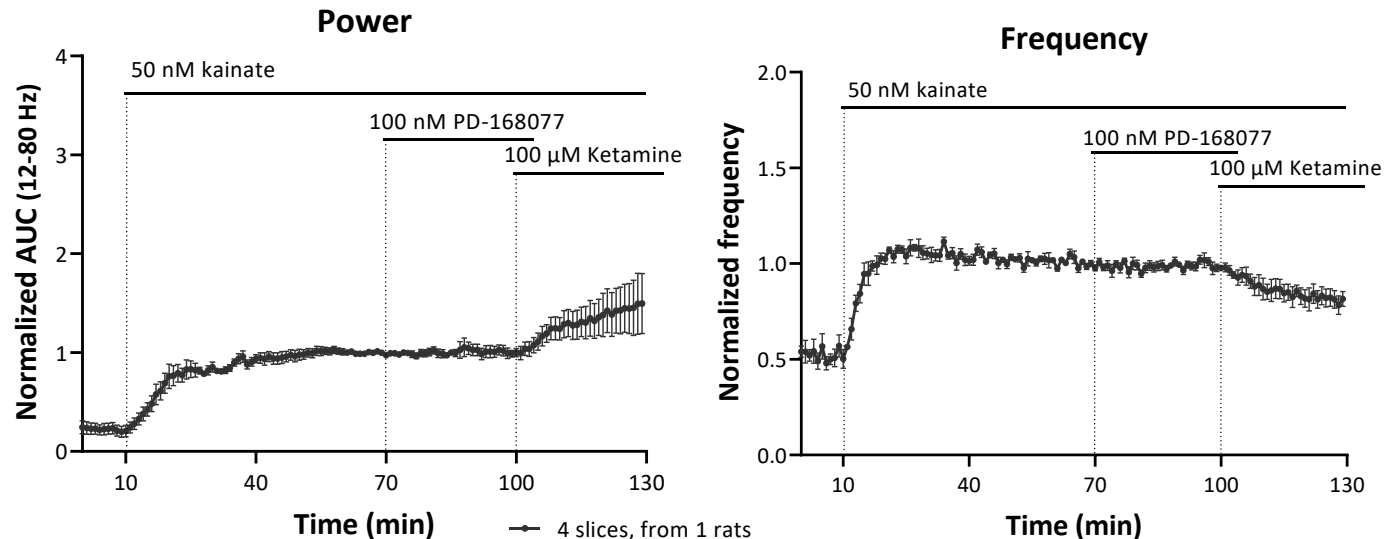**Power**

Frequency

50 nM kainate

10 μM haloperidol

Haloperidol (9 slices, from 7 rats)

Power

Frequency

50 nM kainate

100 nM PD-168077

100 μM Ketamine

4 slices, from 1 rats

Haloperidol
PD-168077

- Haloperidol – a dopaminergic receptor antagonist – applied at 10 μM concentration decreased the power and the frequency of oscillations.

- PD-168077 – a dopaminergic receptor D4 agonist – applied at 100 nM concentration did not modify the power and the frequency of network oscillations. As expected, ketamine applied at 100 μM increased the power and decreased the frequency of network oscillations.
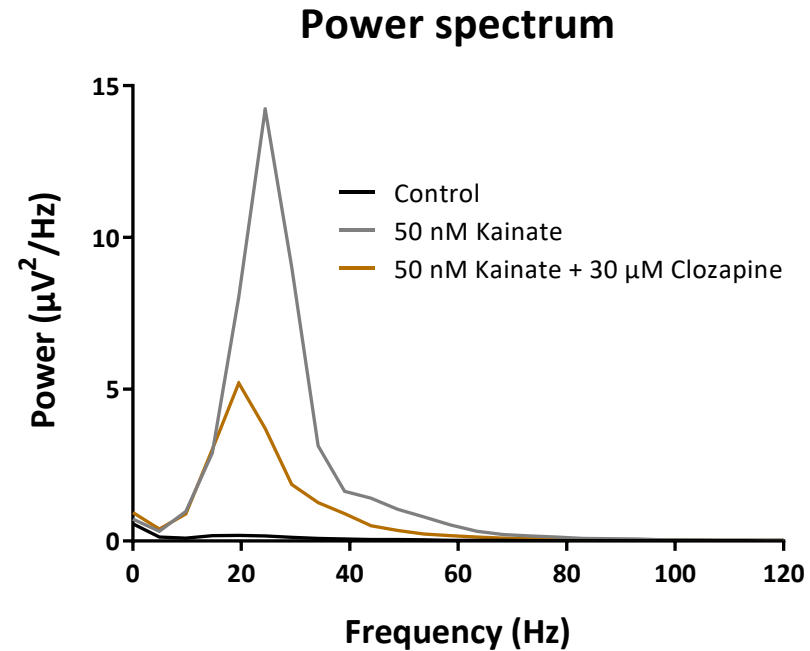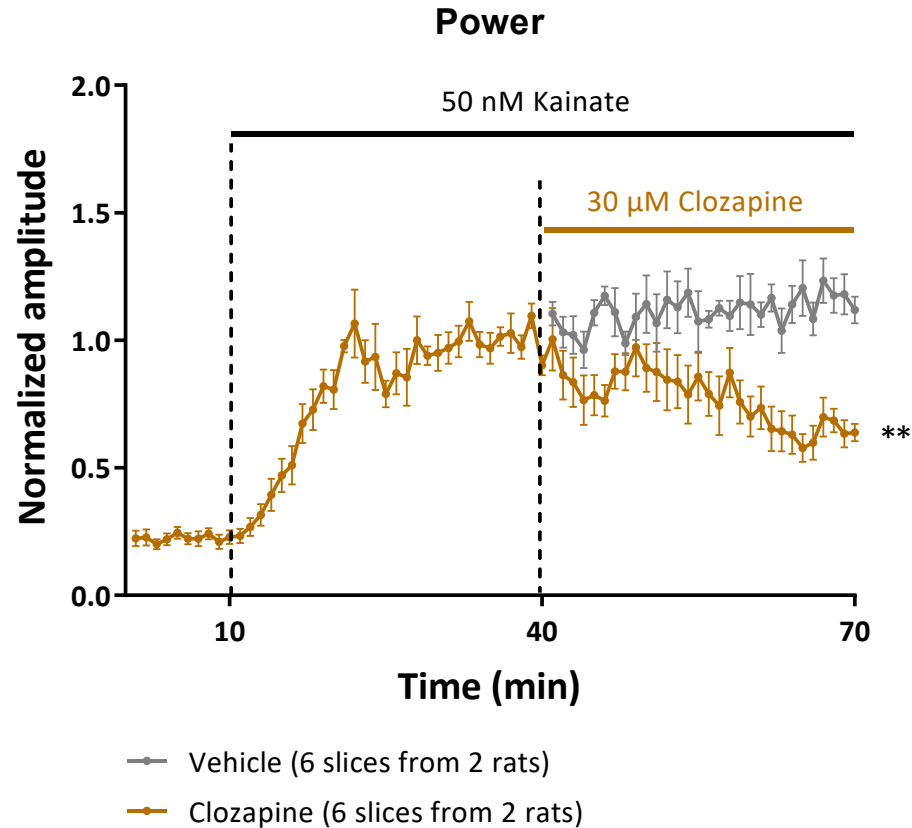
RESULTS

Antipsychotic

Clozapine

Power

50 nM Kainate

30 µM Clozapine

**

— Vehicle (6 slices from 2 rats)
— Clozapine (6 slices from 2 rats)

Power spectrum

— Control
— 50 nM Kainate
— 50 nM Kainate + 30 µM Clozapine

- Clozapine – a serotonin and dopamine receptors antagonist used as antipsychotic agent – applied at 30 µM concentration decreased the power of network oscillations.

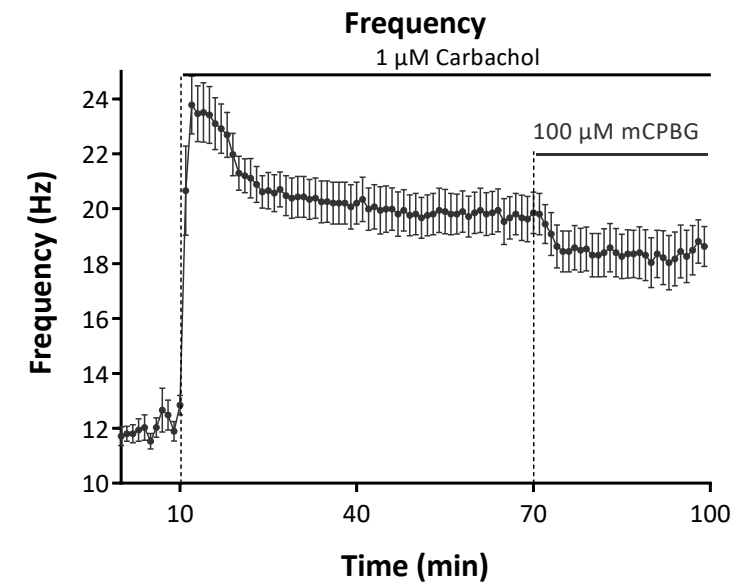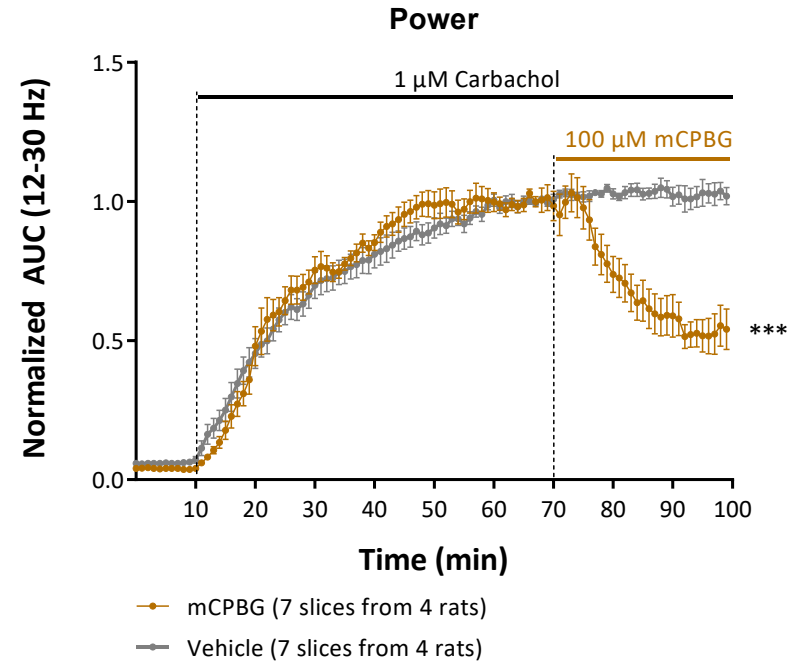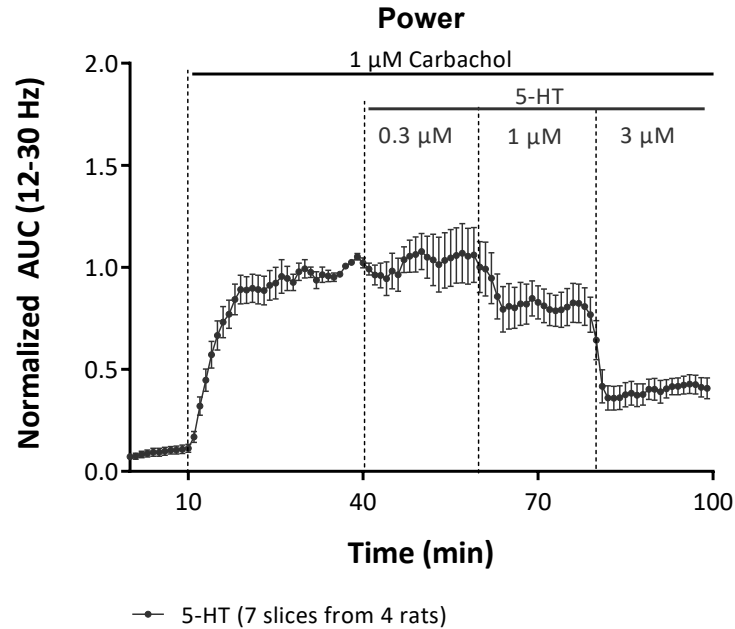neuroservices
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RESULTS

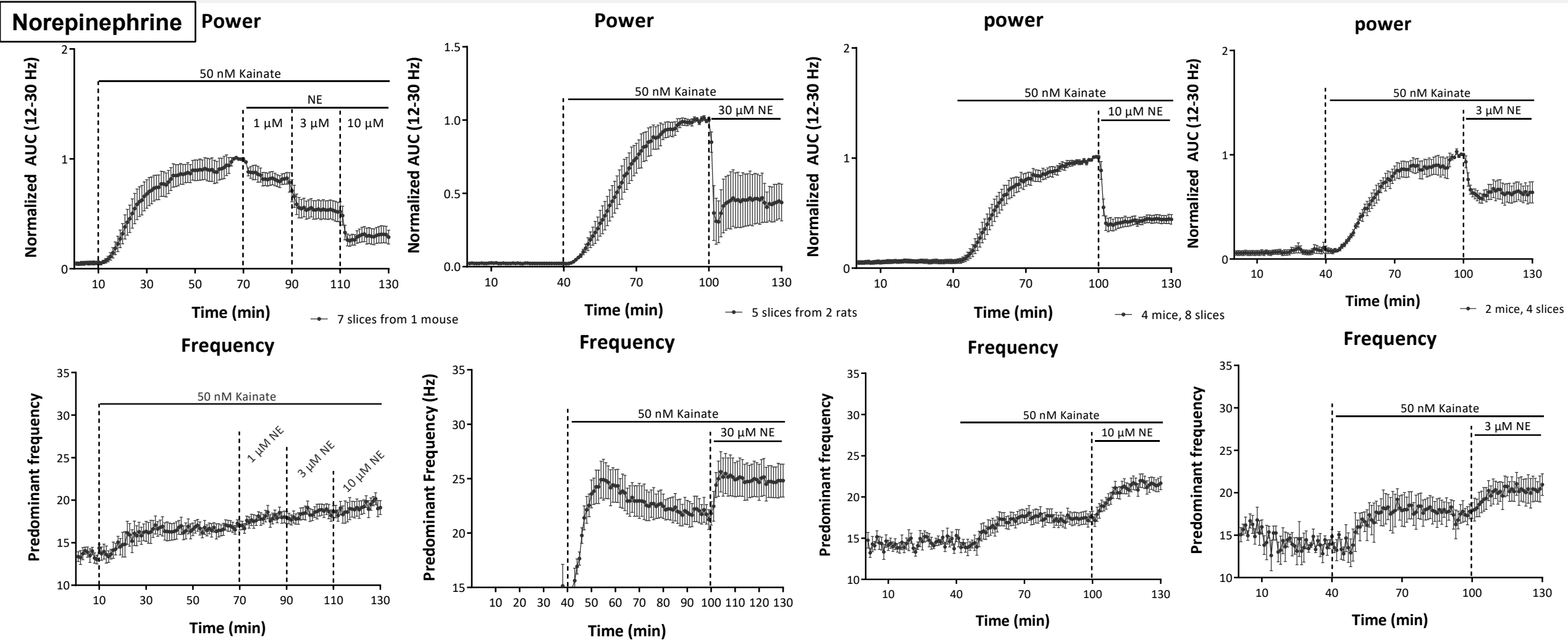## Serotoninergic receptors

5 HT, mCPBG

- 5-HT dose-dependently decreased the power of network oscillations. Similarly, mCPBG – a $5\text{-}HT_3$ agonist – applied at 100 µM concentration decreases the power and the frequency of oscillations .

RESULTS

Adrenergic receptors (1/2)

Norepinephrine

- Norepinephrine (NE) decreased the power and increased the frequency of kainate-induced network oscillations in both rat and mouse hippocampal slices.
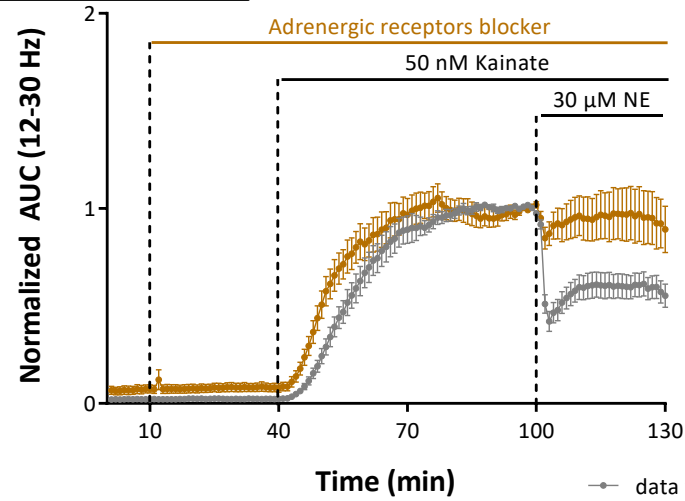
RESULTS

Adrenergic receptors (2/2)

Norepinephrine
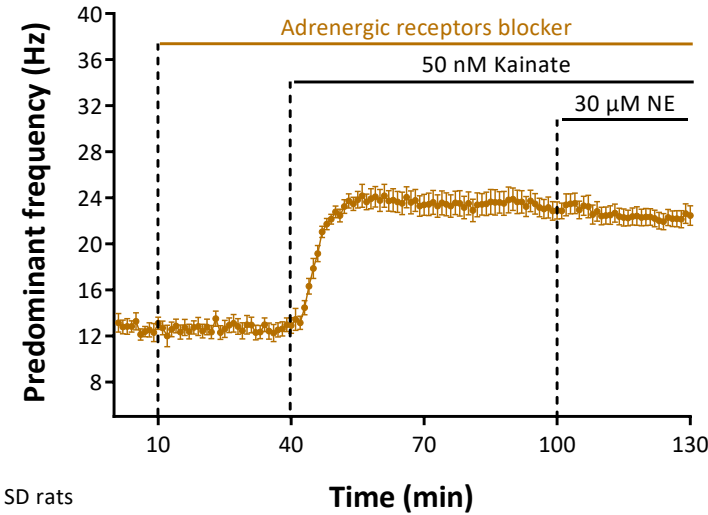
Power

Normalized AUC (12-30 Hz)

Adrenergic receptors blocker

50 nM Kainate

30 µM NE

Time (min)

data from SD rats

Frequency

Predominant frequency (Hz)

Adrenergic receptors blocker

50 nM Kainate

30 µM NE

Time (min)
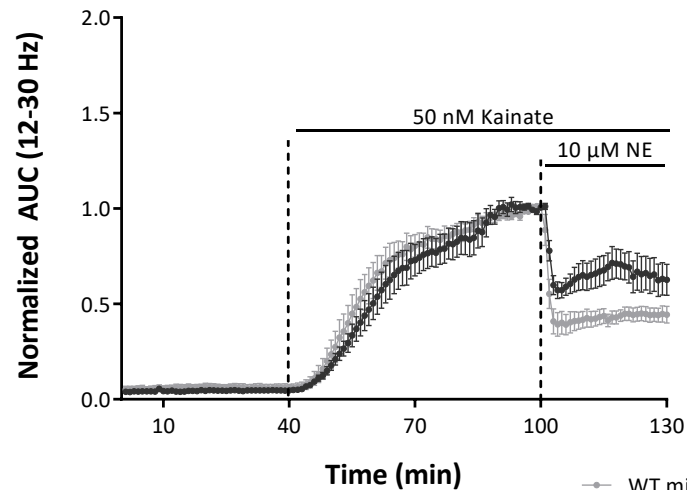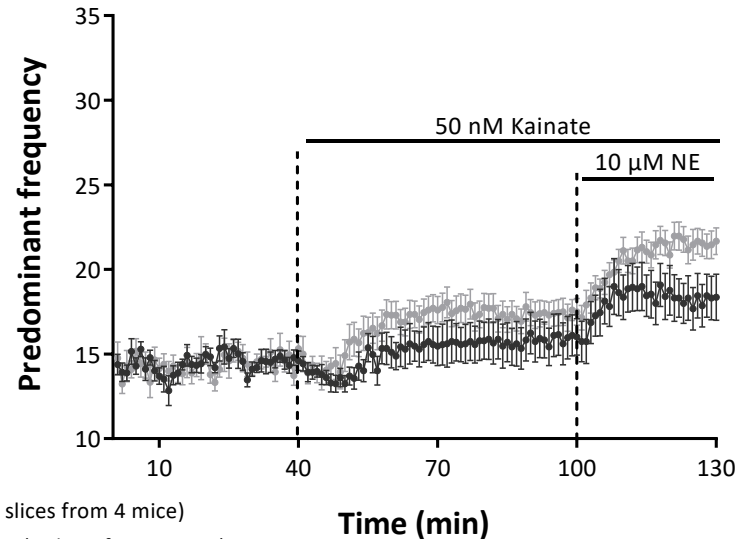
- In the presence of an Adrenergic receptors blocker, norepinephrine did not substantially modulate kainate-induced network oscillations in rat hippocampal slices.

Power

Normalized AUC (12-30 Hz)

50 nM Kainate

10 µM NE

Time (min)

WT mice (8 slices from 4 mice)

$\alpha 1_A$ KO mice (9 slices from 4 mice)

Frequency

Predominant frequency

50 nM Kainate

10 µM NE

Time (min)

- In kainate-induced network oscillations from α1A KO mice, the effect of NE was reduced in comparison with the one in WT mice.

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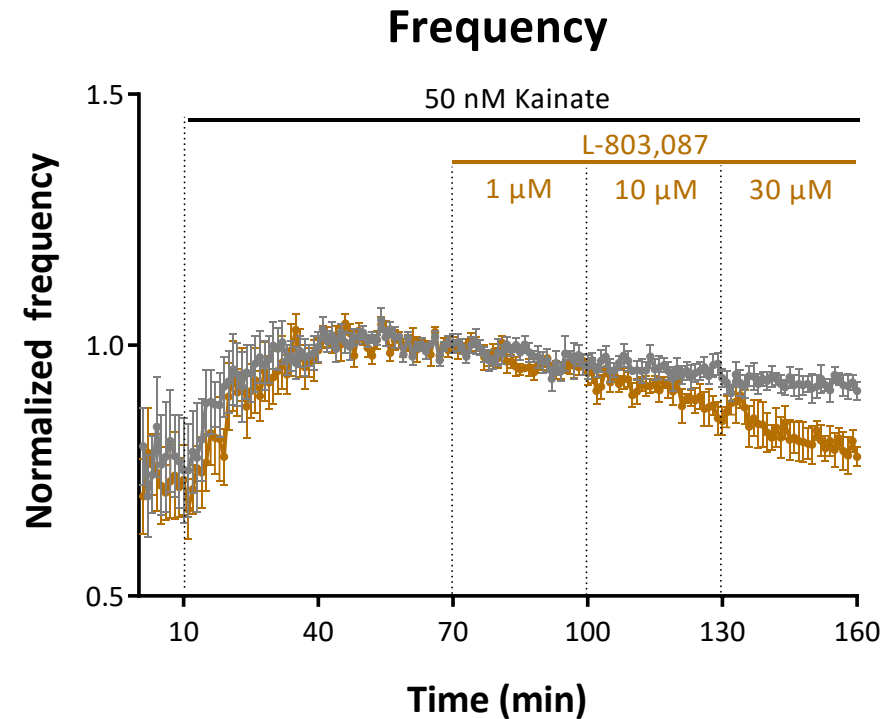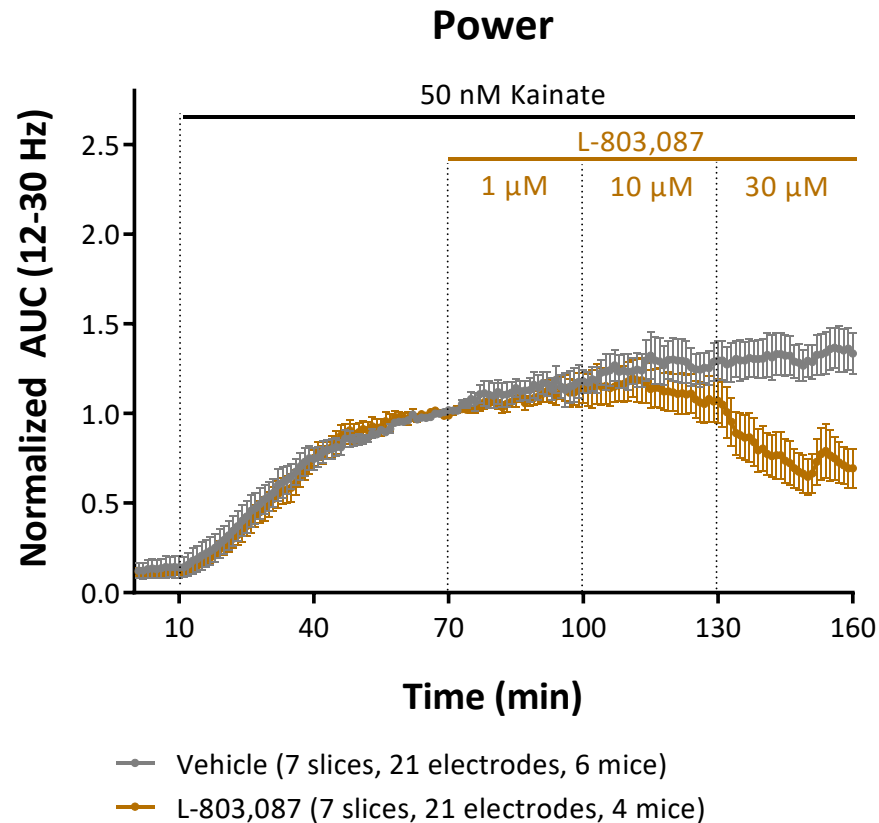ALLIANCE

RESULTS

Somatostatin receptors

L803,087

Power

Frequency

- Vehicle (7 slices, 21 electrodes, 6 mice)
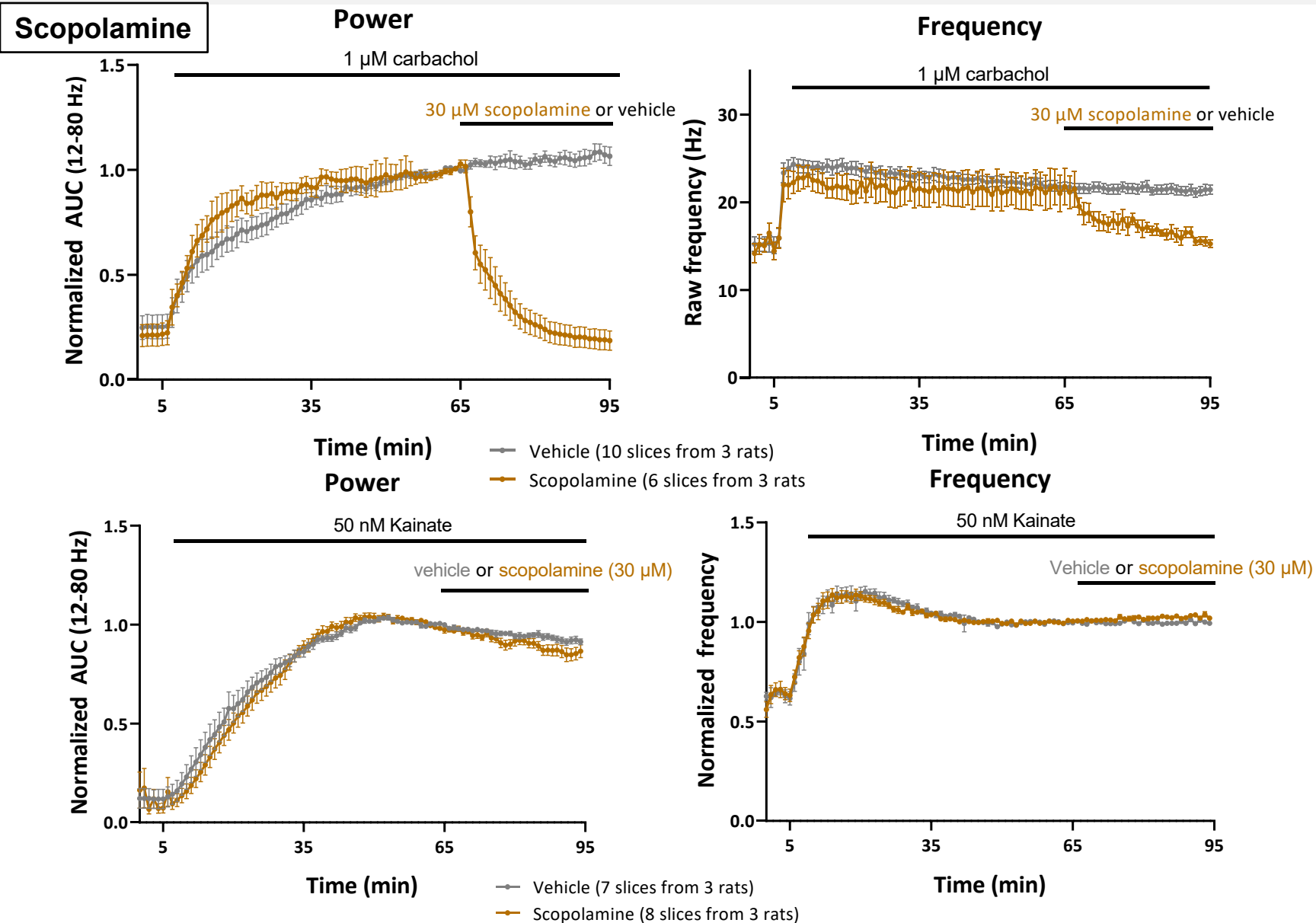- L-803,087 (7 slices, 21 electrodes, 4 mice)

- L-803,087 – a potent and selective SST4 agonis t– decreased both the power and the frequency of network oscillations when applied at 30 µM.

RESULTS

Cholinergic receptors (1/2)

Scopolamine

Power

1 µM carbachol

30 µM scopolamine or vehicle

Normalized AUC (12-80 Hz)

Time (min)

Vehicle (10 slices from 3 rats)
Scopolamine (6 slices from 3 rats

Frequency

1 µM carbachol

30 µM scopolamine or vehicle

Raw frequency (Hz)

Time (min)

Power

50 nM Kainate

vehicle or scopolamine (30 µM)

Normalized AUC (12-80 Hz)

Time (min)

Vehicle (7 slices from 3 rats)
Scopolamine (8 slices from 3 rats)

Frequency

50 nM Kainate

Vehicle or scopolamine (30 µM)

Normalized frequency

Time (min)

- Scopolamine – a non-selective muscarinic antagonist – blocked the network oscillations induced by carbachol but did not modify oscillations induced by kainate.
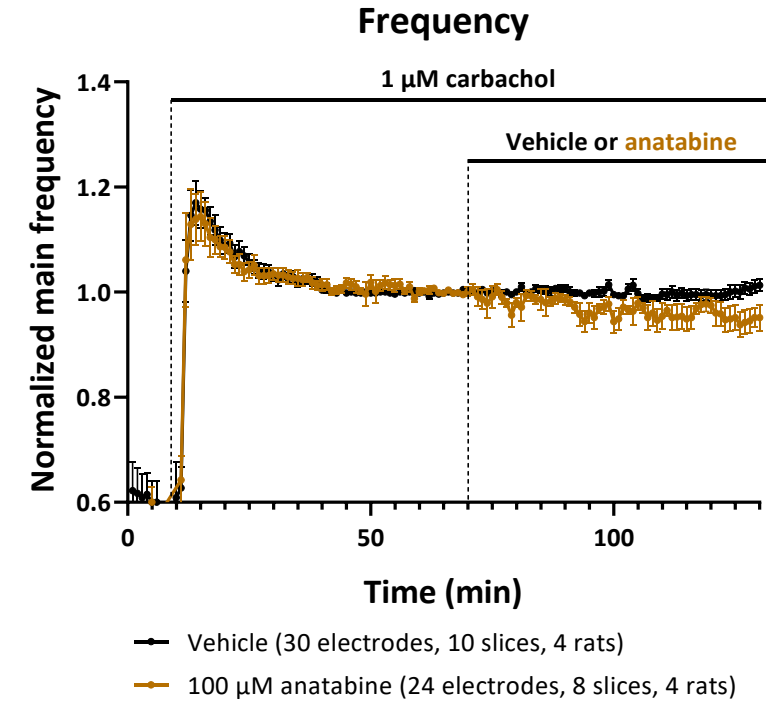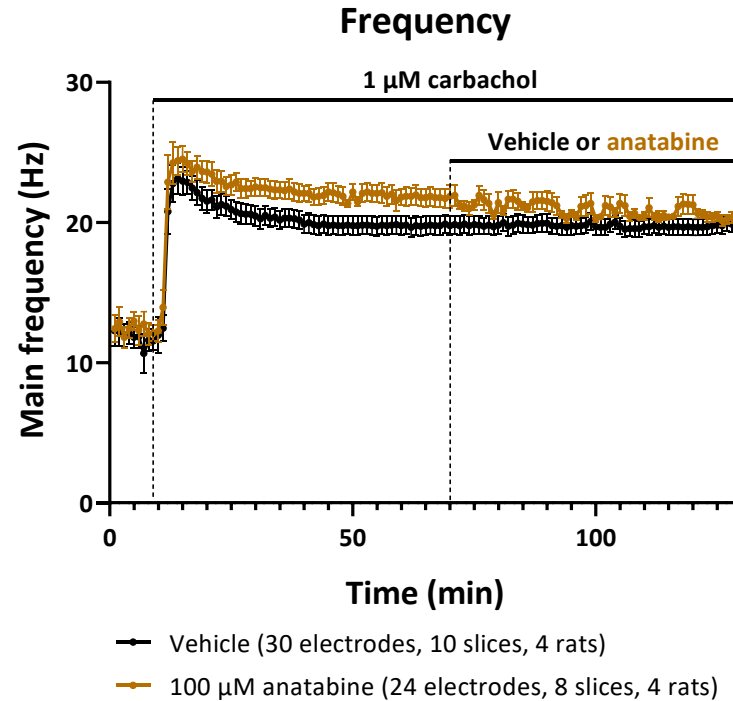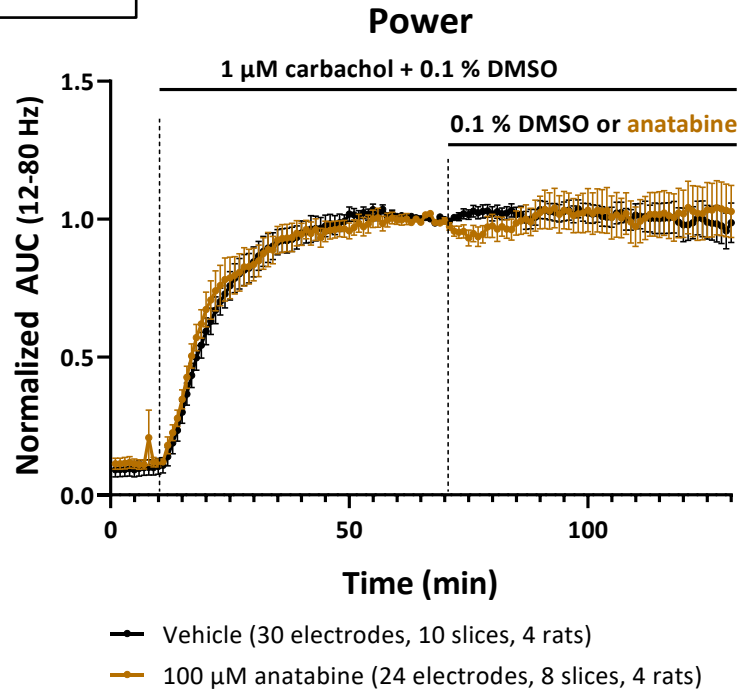
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RESULTS

Cholinergic receptors (2/2)

Anatabine

- Anatabine – a cholinergic agonist – did not modulated the power of carbachol-induced oscillations but did slightly reduced the frequency.
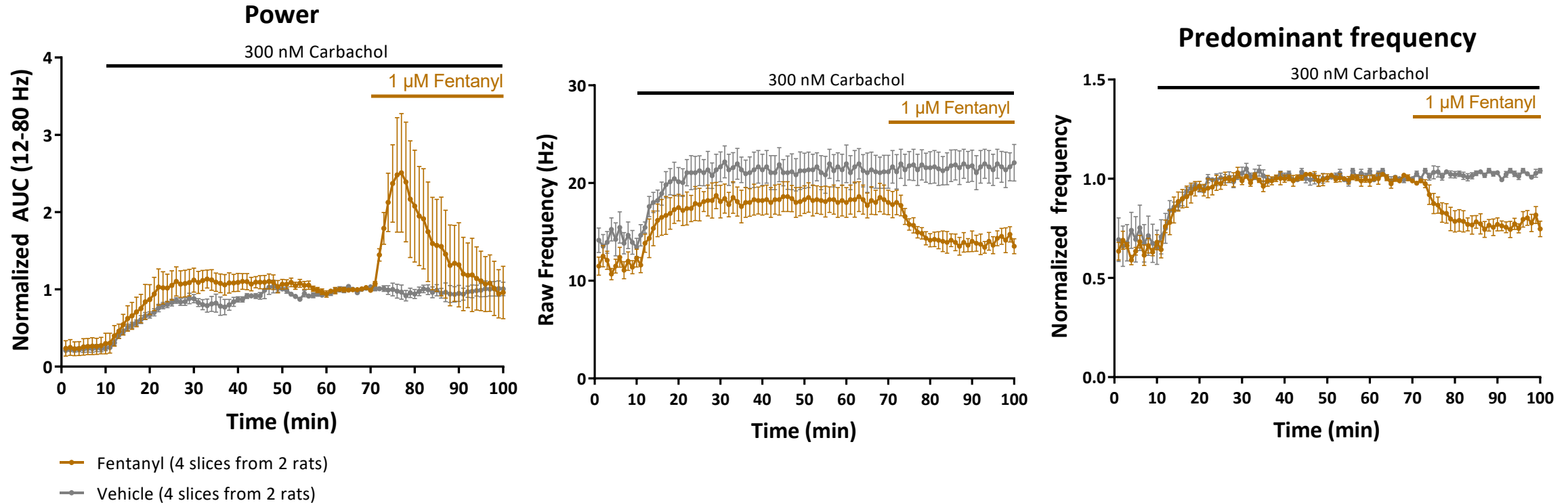
RESULTS

Opioid receptors

Fentanyl

Power

300 nM Carbachol

1 µM Fentanyl

Predominant frequency

300 nM Carbachol

1 µM Fentanyl

300 nM Carbachol

1 µM Fentanyl

— Fentanyl (4 slices from 2 rats)

— Vehicle (4 slices from 2 rats)

- Fentanyl – a µ-opioid agonist – applied at 1 µM modulated carbachol-induced network oscillations in rat hippocampal slices by transiently increasing their power and decreasing their frequency.
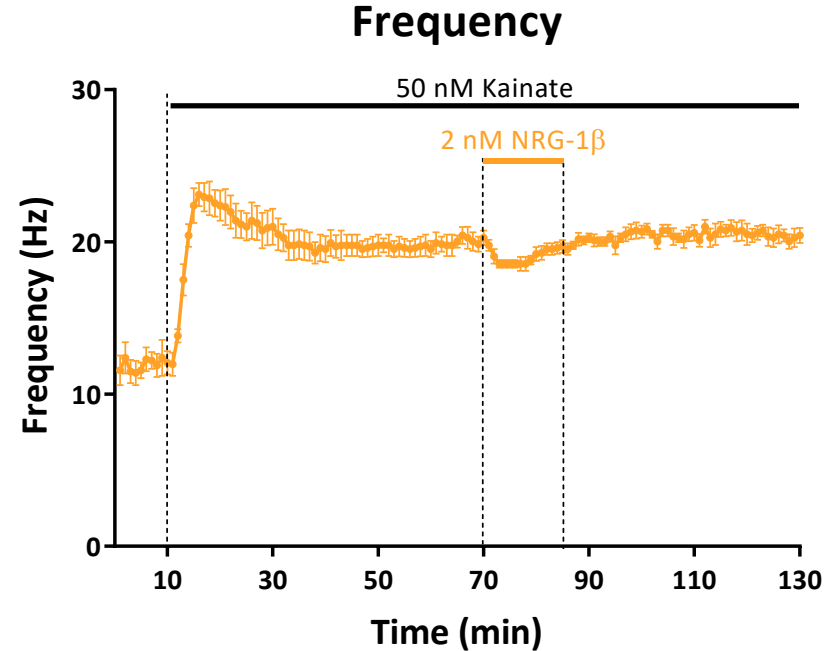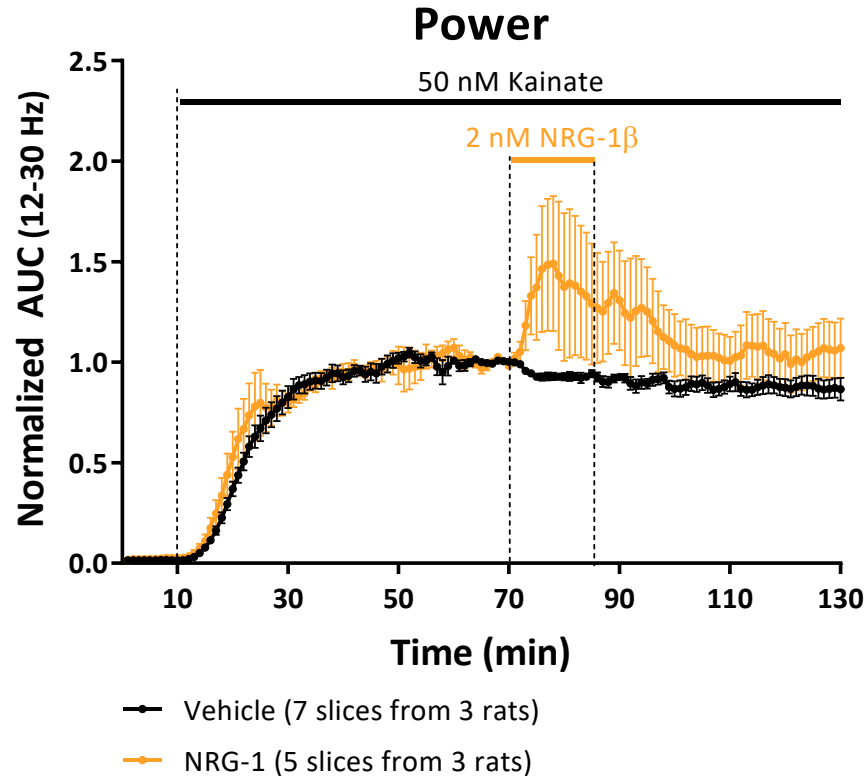
Neuregulin-ERBb4 pathway

NRG-1β

Power

Vehicle (7 slices from 3 rats)

NRG-1 (5 slices from 3 rats)

Frequency

- NRG-1β – an epidermal growth factor receptor (EGFR) agonist – increased the power and decreases the frequency of kainate-induced oscillations.

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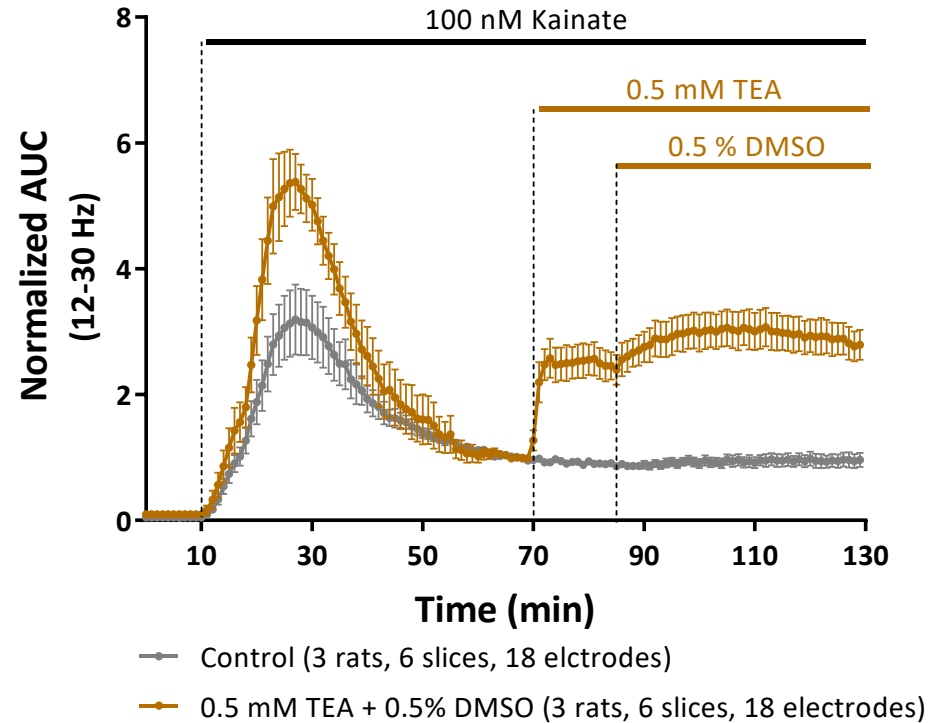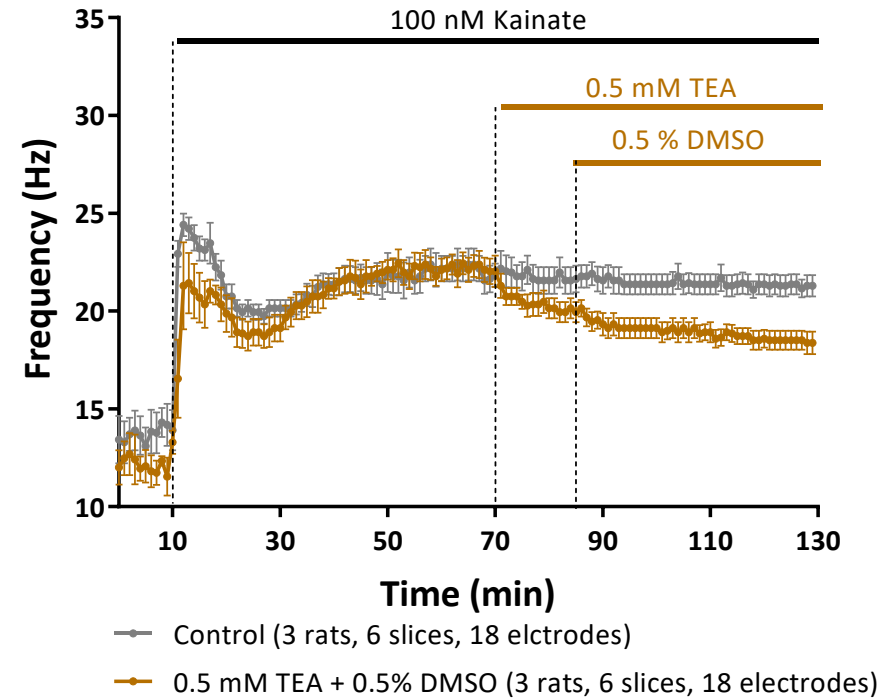A L L I A N C E

RESULTS

SK channels

TEA

Power

Predominant Frequency

- The application of 0.5 mM Tetraethylammonium (TEA) – an inhibitor of Ca^{2+}-activated K^+ (SK) channels - significantly increased the AUC of network oscillations and decreased their frequency.

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